

Durham Research Online

Deposited in DRO:

12 March 2018

Version of attached file:

Accepted Version

Peer-review status of attached file:

Peer-reviewed

Citation for published item:

Girkin, John M. and Carvalho, Mariana Torres (2018) 'The light-sheet microscopy revolution.', *Journal of optics.*, 20 (5). 053002.

Further information on publisher's website:

<https://doi.org/10.1088/2040-8986/aab58a>

Publisher's copyright statement:

Original content from this work may be used under the terms of the Creative Commons Attribution 3.0 licence. Any further distribution of this work must maintain attribution to the author(s) and the title of the work, journal citation and DOI.

Additional information:

Use policy

The full-text may be used and/or reproduced, and given to third parties in any format or medium, without prior permission or charge, for personal research or study, educational, or not-for-profit purposes provided that:

- a full bibliographic reference is made to the original source
- a [link](#) is made to the metadata record in DRO
- the full-text is not changed in any way

The full-text must not be sold in any format or medium without the formal permission of the copyright holders.

Please consult the [full DRO policy](#) for further details.

ACCEPTED MANUSCRIPT • OPEN ACCESS

The light-sheet microscopy revolution

To cite this article before publication: John Girkin *et al* 2018 *J. Opt.* in press <https://doi.org/10.1088/2040-8986/aab58a>

Manuscript version: Accepted Manuscript

Accepted Manuscript is “the version of the article accepted for publication including all changes made as a result of the peer review process, and which may also include the addition to the article by IOP Publishing of a header, an article ID, a cover sheet and/or an ‘Accepted Manuscript’ watermark, but excluding any other editing, typesetting or other changes made by IOP Publishing and/or its licensors”

This Accepted Manuscript is © 2018 IOP Publishing Ltd.

As the Version of Record of this article is going to be / has been published on a gold open access basis under a CC BY 3.0 licence, this Accepted Manuscript is available for reuse under a CC BY 3.0 licence immediately.

Everyone is permitted to use all or part of the original content in this article, provided that they adhere to all the terms of the licence <https://creativecommons.org/licenses/by/3.0>

Although reasonable endeavours have been taken to obtain all necessary permissions from third parties to include their copyrighted content within this article, their full citation and copyright line may not be present in this Accepted Manuscript version. Before using any content from this article, please refer to the Version of Record on IOPscience once published for full citation and copyright details, as permissions may be required. All third party content is fully copyright protected and is not published on a gold open access basis under a CC BY licence, unless that is specifically stated in the figure caption in the Version of Record.

View the [article online](#) for updates and enhancements.

The Light Sheet Microscopy Revolution

J. M. Girkin¹ and M. T. Carvalho²

¹ Biophysical Sciences Institute, Department of Physics, Durham University, Durham, DH1 3LE, UK

² INL - International Iberian Nanotechnology Laboratory, 4715-330, Braga, Portugal

Abstract

This paper reviews the rapid advances that have been made in one form of optical biological imaging in the last decade, namely that of light sheet microscopy. Although the concept was originally presented over one hundred years ago, at the time it was a methodology that lacked the technology to really make it a viable tool for practical everyday imaging in the biologist's laboratory. However, since its re-discovery, it has started to transform *in vivo* and increasingly intact organ imaging in a number of areas of biology. This review looks back at the beginning of the method and then the crucial role that modern optical technology, frequently developed for other fields, has played in advancing the instrumentation. This paper will also look at the Open SPIM route that was developed whereby, through the purchase of a few optical components, researchers have been able to develop their own bespoke instruments and we consider if this may be a route forward for the rapid development of other technological breakthroughs.

1. Introduction

Ever since the use of the first simple magnifying lenses, there has been a desire to image life with increased resolution. The development of more complex multiple lens systems, around four hundred years ago by people such as Robert Hooke, led to the discovery of cells, bacteria, and the plethora of biological activities taking place at levels below the visible resolving power of the human eye. A major desire of the life scientist thus grew to be the observation of life at high spatial resolution and, increasingly, in three dimensions for extended periods of time. The development of the light sheet microscope [1] over the past ten years has been at the forefront in this quest for *in vivo* imaging with minimal perturbation to the sample. The rediscovery of the concept also came at just the right time in terms of technical advances in instrumentation and perhaps also in a wider context, in the way that knowledge is now disseminated and this has helped to advance the field, probably, more rapidly than other new microscopy techniques. As an indication, over 500 references to the technique were found in preparation for this optical review of light sheet microscopy, many in the highest profile journals. This review initially looks at the background and basic optical principles involved in light sheet microscopy, before examining the rapid optical advances that have been made to improve both the quality of the images and the versatility of the method.

The first recorded manifestation of the light sheet microscope was in 1912 when a German chemist, Richard Zsigmondy, and a physicist, Henry Siedentopf, worked together to develop improved methods of understanding colloidal gold. They wished to push the resolution of optical microscopes to observe these nanostructured materials. However, it was already well established that there was a limit to the resolution possible, based on the work by Abbe, and that one needed to use lenses with a high numerical aperture and short wavelengths of light for the maximum resolution possible. Crucially to Zsigmondy and

Siedentopf, an employee with Zeiss at the time realised that, as well as maximising the optical resolution possible, one also needed high contrast images in order to maximise the visibility of any features. They thus introduced the light onto their microscope perpendicular to the observation axis and consequently only saw the light that was scattered by the gold nano-particles into their line of observation [2], in so called Tyndall cones, with the colour being dependent on the size and material of the particle. All observations had to be made by eye as photography had not yet advanced sufficiently to be employed routinely for microscopy, and the light source, although powerful, was variable as it involved focused sunlight. The experiments were recently repeated and further details can be found in an interesting non-biological background paper [3], where the technique is called ultramicroscopy. Consequently, although the method worked well for the inventors and a few future collaborators, also looking at events that could be studied by observation of scattered light such as Brownian motion, and a commercial system was built by Zeiss, the technique was not widely accepted. This was probably due to the in-ability to record the events and the working distances of high NA objectives, at the time was very short making the range of samples that could be viewed limited, mainly to colloidal suspensions.

It was thus about 80 years before the idea of using sheets of light to illuminate the sample was re-discovered. The main focus, then, was on the development of higher resolution imaging methods, and the concept of the light sheet was used to reduce the excitation volume [4–6]. This was followed eight years later by the use of a light sheet with an orthogonal camera to view water borne bacteria [7]. The light sheet here was not particularly thin as it was produced using a simple cylindrical lens with a low numerical aperture. Following these initial publications, many similar methods have surfaced, e.g. OPFOS, SPIM, HR-OPFOS, mSPIM, DSLM, TSLIM [8,9], some of which will be mentioned later in this review. The introduction of the light sheet microscope, as it is now understood, was only described in 2004 [1], which started the rapid growth of the technique. This instrument used the now ubiquitous orthogonally mounted microscope objectives with the light sheet being produced using a cylindrical lens before the illumination objective. With the advent of advanced, and now low-cost CCD and CMOS cameras, linked with the plethora of all solid-state frequency doubled laser sources, covering almost the complete visible spectrum, the technology was now readily available to deliver high performance imaging at a reasonable cost. The optical concept of the light sheet method is shown in figure 1. It was around this time that the acronym SPIM came into widespread use, standing for Selective (or alternatively Single) Plane Illumination Microscopy, becoming a standard term particularly in the field of developmental biology. Thereafter, the acronym LSFM (which stands for Light Sheet Fluorescent Microscopy) was designated to describe all of the techniques based on light sheet illumination.

In addition to the technological development, at this time, the use of genetically encoded fluorescent proteins was also revolutionising the study of *in vivo* biology, the inventors of the method being awarded the 2008 Nobel Prize in Chemistry [10]. This advance, and its implications for the optical methods employed to observe processes within the animal are described later in this review. The other linked biological change was the increasing use of the humble Zebrafish (*Danio rerio*) as a model organism, in particular for developmental studies. As the fish is basically transparent or can be made transparent through the use of pharmaceutical compounds or genetic manipulation [11], optical imaging at depth within a living sample became viable. In their original publication [1] the Stelzer team also demonstrated the use of the technique in imaging developing *Drosophila* embryos and, shortly afterwards, the other main invertebrate model, *C. elegans*, was imaged by multiple groups, as again, it is fundamentally transparent. The method was then taken up by the plant community in particular for the imaging of root development over extended periods of time [12,13].

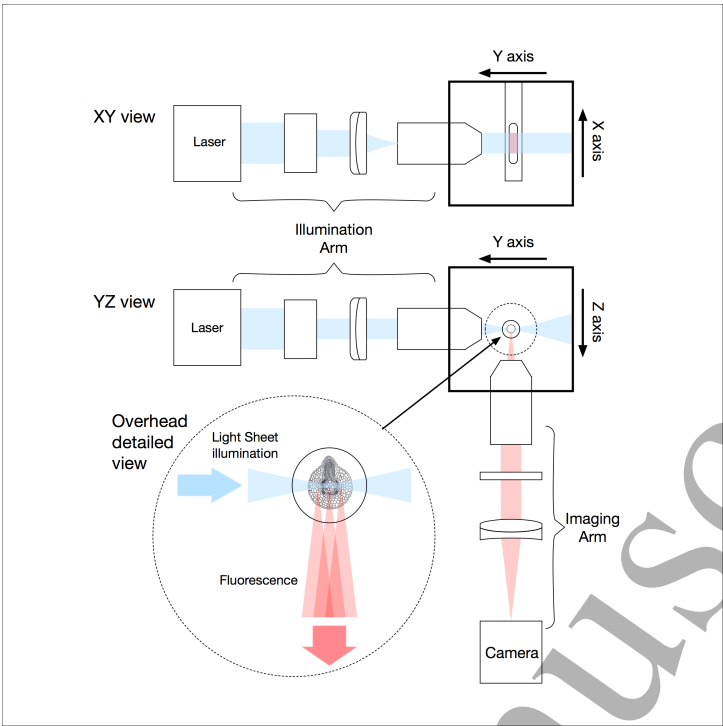


Figure 1. Concept of the LSFM method with the excitation light sheet produced using a cylindrical lens before the excitation objective and the orthogonal imaging arm. The top drawing shows the XY plane view, where the light is kept collimated by the objective unto the sample. The bottom drawing represents the YZ plane view, where the objective strongly focuses the light unto the sample. The addendum (image in the circle), shows in details on the collected fluorescence emitted by the sample.

1.1 Why Light Sheet Microscopy is of such interest to Life Scientist

Before considering the optical and technical background and recent advances in SPIM, it is worth carefully considering why it has so rapidly become the imaging method of choice for significant numbers of research areas, ranging from brain imaging through to the study of heart disease and plant growth. As life can be thought of as a four-dimensional process consisting of three spatial dimensions and time, to truly understand what is taking place within a sample, one needs to observe the system for an extended period of time with minimal perturbation to the sample. Analysis of biological sample dynamics over time, or four-dimensional imaging, represents a major goal of biological science. The ability to resolve volumetric structures, within the cell or tissue, during developmental events, or in response to environmental or experimental stresses, is critical to the understanding of biological behaviour. Within this objective, LSFM has become the tool of choice [14] for many research groups, as SPIM was conceived for *in vivo* 4D imaging.

The conventional method for obtaining three-dimensional data sets in biology is through the use of a confocal microscope [15]. This provides the best possible optical sectioning and the system can be diffraction limited in all three dimensions. However, due to the presence of the pinhole, confocal systems are inherently based on scanning, normally of the imaging beam, and hence are limited to several tens of optical slices per second and, occasionally, up to a hundred optical sections a second in certain home built advanced systems. Therefore, to build up a three-dimensional image, one needs to repeat this optical sectioning at different depths within the sample, leading to volumetric imaging typically taking at least several seconds, and more routinely over 30 seconds in most situations. This

means that three-dimensional dynamic effects are frequently missed. In addition, there is a significant risk of phototoxic events or photo-bleaching of the fluorophore. The excitation light illuminates not just the small volume of interest, but a cone of the sample above and below the focal volume. All of this out of focus excitation is wasted as the fluorescent photons are rejected by the pinhole in the confocal system. However, the excitation has taken place and thus there is a risk of photo-bleaching and other unwanted side effects from the light. So, for long term extended imaging, the method, although offering a very high spatial resolution, is comparatively slow, limited to a typical depth of around 50 microns and has the high risk of photo-bleaching and phototoxic events. The use of non-linear excitation methods does increase the depth of imaging possible and removes out of plane bleaching (or offers no bleaching in the case of Raman or harmonic based methods) but still has to be beam scanned. In addition, the use of longer wavelengths, although reducing unwanted phototoxic events, can increase the risk of localised sample heating and damaging. Non-linear excitation methods (e.g. CARS, multiphoton fluorescence, harmonic imaging) are also inherently more complex in terms of the light sources [16] and significantly more expensive.

The main advantage of light sheet microscopy is that a single optical section is illuminated at a time, and that the fluorescence from all of the molecules excited within that optical section, and within the field of view of the observing lens, are collected. Crucially, only the molecules within the imaging plane are excited, minimising the risk of phototoxic events [17]. The fact that one is collecting an entire optical slice in one go also means that camera-based detection is possible, and beam scanning is not required. This means that optical sections can be recorded very rapidly, enabling transient effects to be observed. Full XYZ high-resolution data sets can be recorded within seconds and, as will be described later, sub-second volumetric imaging is possible [18]. It thus became clear very quickly that, for live imaging, SPIM provided an ideal route to collecting 3D data [8,19]. Since the first publications, there have been multiple reviews written for the different branches of biology, but the overriding message of the advantages of SPIM are 3D speed, minimal perturbation and acceptable resolution, and the limitations are now becoming to be better understood [20]. The optical sectioning method is now being linked with other more conventional widefield microscopy to provide the maximum quantity of data from a single experiment [21] helping to reduce the number of animals used in such research. The technique is also being applied to large specimens [22] as the understanding of the trade-offs between the resolution, the speed of imaging and volume are known.

LSFM imaging, therefore, provides biologists with a new tool to observe processes for longer periods of time in an *in vivo* situation. This review will now go on to look at the basic optical principles behind the method, before looking at the current state of the art techniques, where optical physics is pushing back the boundaries for rapid, deeper, less invasive imaging combined with high resolution. The link between the advances in technology will be closely related to the increased biological understanding that these advances enabled. We also explore areas where there are still technical challenges and where new optical methods are required to overcome the biologically imposed barriers.

2. Principals and Practical Implementation of Light Sheet Microscopy

The core concept behind the light sheet microscope is very simple, and shown in figure 1. A sheet of light is produced and illuminates the sample, which in most biological systems is fluorescence, which is emitted in all directions, is collected using a second microscope objective, which is perpendicular to the excitation sheet, thus capturing an entire optical slice [1,23]. The collected light is then re-imaged onto a camera and, typically, the sample is moved through the light sheet to collect the next optical plane. The crucial step is in producing a uniform light sheet which is coincident with the focus of the observing lens.

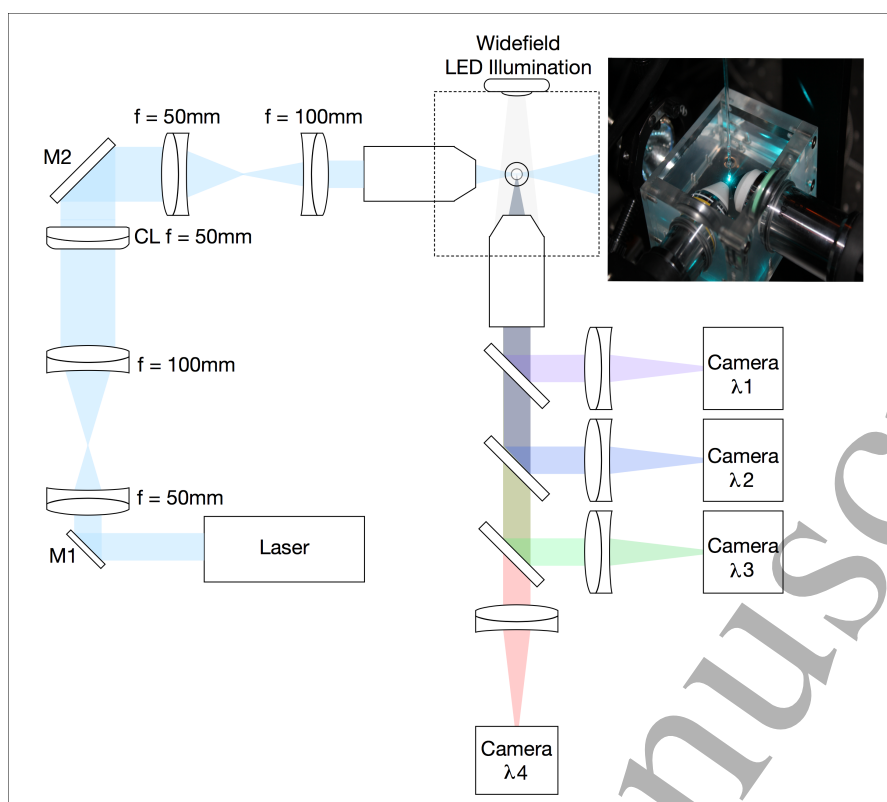


Figure 2. Practical multi-wavelength LSFM system with typical optical components and including widefield LED illumination (not drawn to scale).

In figure 2, a practical implementation is shown, where light from a suitable laser source is expanded and directed through a cylindrical lens, and the resulting line of light being re-imaged into the back aperture of an objective lens. This line of light fills the full aperture of the illumination objective in one axis, leading to the full focusing power of the lens (determined by its numerical aperture (NA)). However, in the other axis, as only a very small portion of the back aperture is illuminated, the effective NA of the objective is now very small and hence the light is not focused. This leads to the formation of a sheet of light at the focus in which its vertical height is mainly determined by the power of the cylindrical lens, and the thickness of the sheet, by the NA of the objective. Typical lens values are shown in figure 2, and in our home built system the excitation objective has a magnification of 10 times, NA of 0.30 and working distance in water of 3.5 mm (Nikon). This produces a light sheet of around 2 microns' thickness over a field of view of around 150 microns. Clearly here, there is a compromise in terms of the light sheet, if a lens with a higher NA is used, one will have a thinner light sheet, however, the light sheet will not be as uniform across the full field of view of the observation objective. As will be discussed later, optical changes to the illumination source has been one of the areas in which optical advances have been made for improved imaging. Greater details on the basic optical system and considerations given can be found in an earlier paper by Swoger and Stelzer [24] and some advance considerations of the use of a cylindrical lens in this configuration have also been reported [25]. The former paper also describes some of the more detailed considerations of the control hardware and selection of specific components. Variations on this basic illumination configuration can be found later in this review.

The light has now reached the sample and a crucial consideration here is exactly how the sample is mounted. We are generally dealing with live and developing samples and, although normally anaesthetised, they are still subject to movement. As the objectives

typically used are water immersion lenses, the sample is normally mounted in low melting point agarose or water within a glass or Fluorinated Ethylene Propylene (FEP) tube. The advantage of the FEP tubing is that it is approximately refractive index matched to the surrounding water, the disadvantage is that it is not optically perfect and thus can lead to an increase in scattered excitation light [26]. An alternative is to directly extrude the sample in the agar. All methods work and it is frequently down to a user's personal preference or sample. The sample needs to be able to be moved in the Z direction (through the light sheet) in order to record the images at different depths. Due to the complexity of having to keep both the light sheet position and imaging focus in the same plane, moving lenses is not the normal preferred way of depth scanning. Rotation of the sample is also a useful consideration as this helps with the alignment of the correct view into the camera. If this is automated it also means that the same system can be used for a variation of optical projection tomography [27].

Having now correctly illuminated the sample, the critical factor in any optical microscope is the collection of the precious fluorescent photons. The optical properties, of the collection objective, determines the ultimate resolution of the system. Typically, as in our home built SPIM, systems use a water immersion objective with a NA of around 0.8 and a working distance of 3.0 mm and x16 magnification (Nikon). The important practical considerations are the NA and the working distance (WD). If the WD is too short, the lens will hit the excitation objective before it is focused in the light sheet. The collected light is then re-imaged onto a suitable camera, or in the case of systems to be used with multiple fluorophores, a series of dichroic beam splitters can be used to a multiplicity of cameras. A useful addition to this basic configuration is an LED mounted behind the sample which can provide a wide field image which is very helpful when trying to first align a sample where a single optical slice can be difficult to interpret, especially in a sample with only certain cells, or organs labelled. This basic configuration has, however, be subject to a large number of variations, which are discussed below.

2.1. Improvements on Basic Illumination

The illumination system on a light sheet microscope is probably the most important consideration, and variable, as it provides the optical sectioning in the method and also the evenness of illumination across the sample. Consequently, there has been considerable interest either improving the illumination or, more normally, optimising it for a specific application [28] and the range of different methods is now very large. Unless exotic beam forms are used (Bessel and Airy beams are discussed later), the main target has been to reduce sample scattering and absorption from affecting the image quality. The absorption when using both plants and fish can be a major issue, though in the case of fish, as mentioned above, this can be reduced either through the use of the "Casper" no melanocyte fish [11] or with chemical agents to reduce melanin production, in addition to other tissue clearing techniques [29]. If the light sheet entering the sample does impinge on an absorption site it means less light at the focus and hence dimmer images in certain areas, creating regions of shadow in the image. Scattering also leads to a loss of fluorescence excitation at the focus, but the situation can be made worse by the scattered photons causing fluorescence outside the focal volume leading to a loss of contrast in the final image.

The first route to improving the excitation was to introduce excitation light from counter propagating directions such that the beams were fully overlapped in the centre of the imaging lens [30], named as mSPIM for multiple beam SPIM. Whilst this does significantly reduce the shadowing effect on the final image, optimisation is a challenge as one has to align two-counter propagating beams perfectly coplanar to the focal region of the observation lens, looking normally to the excitation. Besides taking two consecutive images,

illuminating the sample from each side, the method presented in this paper reduces the shadowing effect by pivoting the light sheet within the focal plane of the detection optics, thereby illuminating the sample from a range of angles during the camera exposure, obtaining a more even illumination. This concept was later enhanced as a method [31], where a thin pencil of light (a conventionally focused light beam) is scanned rapidly in the vertical direction without the use of the cylindrical lens. Using a resonant scanner, the beam moves rapidly across the field of view such that on a single camera exposure it appears to provide an even light sheet of illumination, it was termed DSLM, for digital scanned laser light sheet fluorescence microscopy, or also d-SPIM for digital SPIM. In both of these beam-scanned cases, the quality of the images is improved, but at a significantly increased complexity in the case of the multiple beam. The Zeiss commercial Z1 instrument does use the beam scanned option (with a cylindrical lens) and the opportunity to have a bi-directional light sheet. Within our laboratory, we have included a scanning mirror (Fig 2, M1) which can be activated if required by the sample, as in ref [30], but we do not illuminate the sample from both sides. For most applications, we have found that this works well enough for samples that are affecting the incoming excitation beam.

A further variation on the excitation method, but still using Gaussian focusing beams, is through the use of so called “HiLo” illumination imaging [32]. Here two images are recorded, the first with the scanned beam, as in DSLM described above, and the second in which the line is turned on and off to produce a grid illumination on the sample (a series of excited and non-excited lines). The two images are then combined using an algorithm utilising the high (Hi) and low (Lo) spatial frequency information from the two images. This method has the advantage of improving the optical sectioning by helping to remove scattered light as the scattered light losses the spatially encoded information from the structured illumination. Inherently the imaging processes takes twice as long and the images require post processing and this can become a major issue with the extended imaging of samples as the data sets can become very large if the raw images are saved.

2.2. Detection Improvements

Light sheet microscopy differs from nearly all other forms of optical microscopy as the excitation and collection optical paths are orthogonal. This opens up new opportunities to improve the quality of the recorded image, without interfering with the illumination light path. Since the first SPIM microscopes, the largest single advance has been in the detectors. The initial SPIM system, in 2004, used a conventional CCD camera but variations have now been built using EMCCDs [33] and high speed cameras for calcium imaging and particle tracking. As with all optical microscopy methods, light sheet systems do not operate well in samples which scatter the light leading both to a loss of signal and an increase in background noise which contributes to the overall loss of contrast. With sCMOS cameras, however, it is possible to only expose a line of the detector pixels effectively producing a confocal line detector system. In one development, this line scanning of the detector was synchronised to the lines scanning of the light source, such that the detection line was imaging the beam scanning through the sample [34]. This use of the “rolling shutter” on the camera removed light from outside the direct line of excitation and improved the quality of the images seen at depth within the sample. Before the advent of the sCMOS camera such a technique would have been possible, but slow and complex using a mechanically controlled slit across the detector.

In a variation of the mSPIM two-beam excitation mentioned above and this concept of the rolling shutter, two imaging objectives have been introduced, so that the sample is both viewed from two sides as well as being illuminated from two directions [35]. The combination again led to an improvement in image quality with the reduction of light scattering and also high speed imaging as the two light source line scans were interlaced,

such that one camera was exposing whilst the other line scan was moving. The disadvantage being the complexity in both the optical configuration, one now has to align the focal volume of four microscope objectives, and also the timing between the different light sources, the movement of the scanning beam and the camera rolling shutter which meant that a dedicated FPGA was required to control this MuVi-SPIM [36]. In addition, to maximise the benefit and to ensure excellent image alignment from the cameras, a significant level of post processing was required. The benefit was improved images at twice the rate of volume imaging as in effect the time normally “wasted” while the beam was moving and the camera preparing the next shutter exposure is spent recording an image with the other camera.

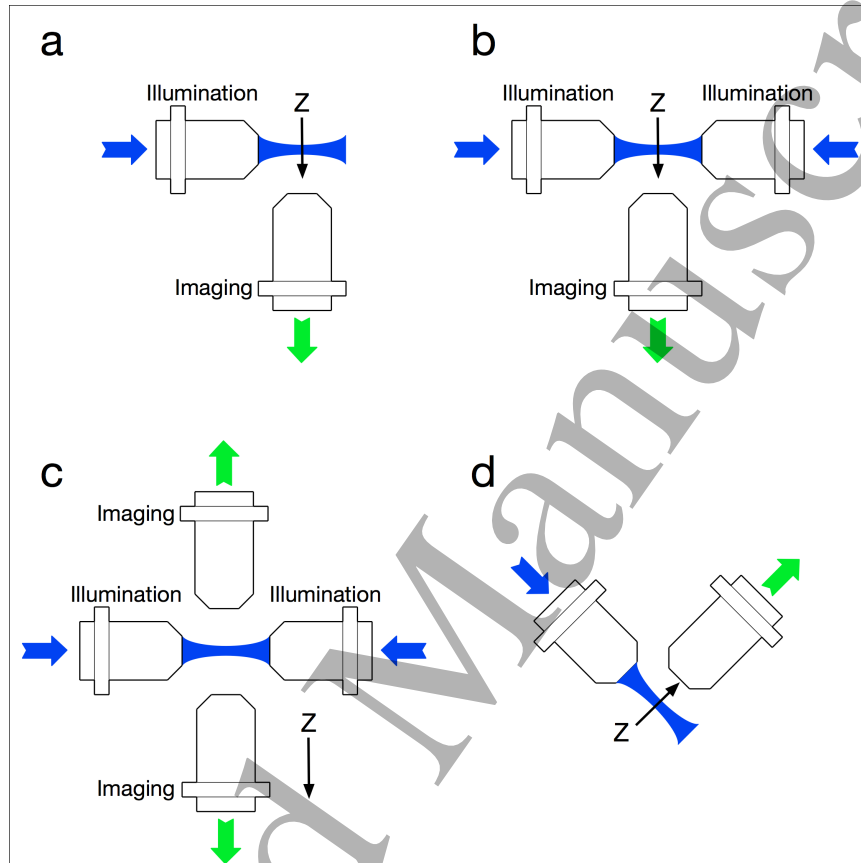


Figure 3. Standard LSFM configurations. (a) SPIM with a single illumination and orthogonal imaging arm, (b) mSPIM with bidirectional illumination, (c) MuVi-SPIM with bidirectional illumination and imaging and (d) iSPIM with a single illumination and orthogonal imaging arm but in an inverted position.

The other technique that has been applied to improve the quality of the images, taken at depth within the sample, is that of adaptive optics. In order to take images more deeply within the sample, one has to look through the tissue, and this induces both scattering, where the above method can improve the images, and aberration of the light, hence the image gets distorted. The simple analogy is looking through a window in which there is a raindrop, which gives you a localised distorted view of the sample. Adaptive optics have been applied before for biological microscopes [37] and showed significant improvement for imaging deeply within samples [38,39]. Crucially as well as correcting for sample induced aberrations the method also removes aberrations that may be present within the imaging optical path. Such aberrations may arise from the method of holding the sample as well as slightly miss-aligned optics or optical elements that are not perfectly corrected. In the case of light sheet microscopy, the use of adaptive optics in the imaging

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

arm of the microscope significant helped to “flatten” the image when the sample was held in a glass pipette and also improved the image quality, allowing to imaging up to 500 microns into the sample [40] with high resolution and contrast and it has also been demonstrated that in a live beating heart the aberrations detected are linked to the heart movement [41] causing loss of resolution as you image through the sample. As light sheet microscopy moves towards larger, and hence thicker samples, these improved methods of detection are becoming increasingly important. Figure 3 illustrates the basic SPIM configurations and nomenclature that has become accepted in the field.

2.3 Open Source systems and Software

One unique feature with light sheet microscopy is the OpenSpim option which was probably the first truly open access high performance scientific instrument [42–44]. Although image processing software had been readily available as open access (ImageJ and Fiji) the complete plans for building a microscope had not been released in this manner. The associated web site provides all of the component lists and assembly drawings required to build you own SPIM system. There are even the designs for 3D printing certain components including for example the sample holding system. This documentation has now been further supported by on-line videos that show people building a system and providing useful tips on how to obtain the best performance from the system. As well as enabling an advanced imaging method to be built at a reasonable cost, within a user’s laboratory it also provided a stimulation for a life scientist to develop collaborations with optical physicists and instrument builders. Although the assembly is not complex for an experienced optical instrument researcher, it is harder for those life scientists who had previously only used commercial microscopes. Thus, life scientists have spoken with optical experts within their research organisations to collaborate to build an OpenSpim system. These collaborations have frequently formed the focus of a final year undergraduate project but more importantly have open up the potential for further interdisciplinary collaborations. Many of the papers outlined later are built on the OpenSpim chassis with novel optical improvements to solve challenges posed by specific life science investigations.

This model is only really viable when the control software is also released, and early versions were released linked with the Micromanager open source package with an option to run on an Arduino computer board if the user so desired [45]. Variations on this have also been developed and reviewed enabling more complex experiments to be undertaken [46]. In light sheet microscopy, many of the challenges really start once the system is ready to run and the data has to be captured. As one main feature of the technique is the speed and long term imaging potential of the system, datasets can become very large very quickly and generating greater than 2Tb of data from a single imaging session (24 hours) is not unusual. This means that new ways need to be considered at the start of any experiment, for both the post processing of the data [47], the storage and the normally simple task of taking the data from the laboratory to the computer on which you may do the image processing. Further details on the way this is handled in the OpenSpim configuration has been described [48] and simple 3D reconstruction is now implemented as Fiji plugin to correlate multiview 3D images [49]. The full challenges of the data processing are beyond the scope of this review paper, which focuses on the optical aspects of the method. To give an indication of the current routes being investigated, ClearVolume is an open-source package for live graphical processing unit (GPU)-accelerated 3D+t multichannel visualisation [50] specifically designed to deal with the large 3D and time data sets. As mentioned above there are now multiple variations on the OpenSpim core instrument and these have been reviewed where their different features are compared (DIY-SPIM, OpenSPIM and OpenSPIN) [51].

3. Genetic manipulation and SPIM Applications

Before considering the recent optical improvement being made in SPIM it is important to appreciate the biological questions of interest and the associated biological innovations. Recent years have seen a number of such technological innovations, which are opening up totally new ways to study the development of complex multicellular organisms. Novel approaches to bio-imaging and automated computational image processing allow the design of truly quantitative studies in developmental biology. The challenge is to understand how an embryo's genes, proteins, and cells, function and interact to manage morphogenesis, cell fate specification, and patterning. With the new advances in biological imaging described above, it is possible to: track morphogenesis and mitotic lineages, visualise dynamic gene expression, cell signalling, optogenetic control, cell behaviour, cell fate choices, cell interactions during tissue growth. As they may be analysed *in vivo* for entire organisms and throughout embryonic development [52].

The use of genetically encoded fluorescent proteins changed the study of *in vivo* biology [10] enabling the transition from single organs to organisms, and from images of fixed, stained tissue sections to movies of *in vivo* embryogenesis. Actually, single cells, tissues, whole organs, and entire embryos can be specifically marked with genetically encoded proteins of various colours [53]. Once a protein is genetically identified, it can be manipulated *in vivo* or *in vitro*, so a specific cell produces these fluorescent molecules that can be identifiable through optical fluorescent imaging. Fluorescent markers can also be manipulated to attach to specific sites (organelle, membrane, nuclei) in the cell. *In Vivo* manipulation can be performed by breeding transgenic animals. The transgenic manipulation can occur in all the cell of the organism, or in specific sites, making use of organ-specific promoters, which involves protein over expression or protein knockout from within a short gene sequence. The transgenic manipulation can also be produced using drug-induced determination processes that can be either total or organ-specific.

Light sheet-based techniques are a powerful tool for live imaging of biological specimens, as they provide an exceptionally high imaging speed, high signal-to-noise ratio, low level of photo-bleaching and good optical penetration depth [54,55]. This unique combination of capabilities makes light sheet-based microscopes highly suitable for live imaging applications, facilitating long-term observation of living biological processes. LSFM are particularly well suited for fast and long-term imaging of developmental processes at multiple spatial and temporal scales. Its application goes from imaging gene expression patterns and molecular interactions to the reconstructions of early developmental dynamics and *in toto* imaging of large fixed specimens [36,56,57].

Within only a few years, light sheet microscopy has contributed substantially to the emerging field of real-time developmental biology. High-speed multiview acquisition and low photo-toxicity have made SPIM a popular choice for studies of organ morphogenesis and function in zebrafish, *Drosophila*, and other model organisms. Applications include: from genetically encoded calcium indicator [58–61] allowing the acquisition of functional imaging [58,60] and FRET process [61], to genetically modified embryos, expressing a variety of fluorescent proteins. As an example, the zebrafish [11] can be *in vivo* manipulated to create genetically modified families, as the transgenic lines (NpHR-mCherry), Tg(kdrl:GFP), Tg(myf7:DsRed), and Tg(myf7:GFP), all now commonly imaged using SPIM [62–66].

The variety of biological applications and the current diversity of light sheet-based techniques illustrate not only the growing popularity of the technique but also suggests a bright future ahead. LSFM has transform *in vivo* and intact organ imaging in a number of areas of biology, as it is much faster and gentler than other established fluorescence microscopy techniques [67]. Fundamental challenges for live imaging proves that SPIM is an indispensable tool for the *in vivo* long-term imaging of large developing organisms, as in

zebrafish embryogenesis [68], and investigations of cellular and sub-cellular dynamic processes [69], as in cell division dynamics [70]. Driven by technical advancements, it is expected that more demanding biological questions awaken. Over the up and coming imaging performance of the microscope we can expect powerful add-ons, for example, photo-activation [66], laser ablation [71], and fluorescence correlation spectroscopy [72], which will be discussed later. Biological research will look deeper into established and novel model organisms and, for the first time, will focus on rapid intracellular processes and morphogenetic movements of tissues during the course of hours [14].

4. Recent Optical Advances in SPIM

The liberty of controlling the illumination arm and the detection arm separately, has given researchers many ideas on how to adapt and change the original LSFM techniques [1,4,5] to fit specific needs predominantly in the excitation path. The major reason for this is that in terms of the detection there are few direct improvements that can be made beyond multiple cameras [35], higher performance cameras with specialised features such as the rolling shutter [34] and the use of adaptive optics to remove induced aberrations from the sample and system [40]. In the multidirectional illumination route, there are the mSPIM, MuVi-SPIM [36] and SIMView-SPIM [73], combining multiple illumination directions and multiple view directions to create a Multiview [35] image of the sample, and these modifications were discussed above. The next increase in complexity comes with IsoView [74] where, with the addition of extra optics and two more cameras, each arm can be used for illumination and detection sequentially or simultaneously.

The first major advance beyond those already described, which mainly consist of using multiple beams, was the introduction of structure encoded onto the beam pattern [75,76]. In the first of these publications the conventional method of structured illumination was used implemented on the SPIM system using a digitally scanned light sheet, but in this configuration the excitation laser was passed through an AOTF (acousto-optical tunable filter) before being digitally scanned. The AOTF placed a sinusoidal intensity pattern onto the excitation light as it was scanned, such that the resulting time averaged intensity pattern for a single camera shot had a sinusoidal intensity pattern across the light sheet. In line with normal structured illumination, the phase of the pattern was altered at to give three images at 0, $2\pi/3$ and $4\pi/3$, and the sum of the squares of the difference of the images was calculated. This process meant that only light emanating from the combined focal plane of the light sheet and observing lens was visualised, removing the fluorescence generated out of focus, thus the scattered light. This enabled the imaging of *Drosophila* embryos for over 58 hours at high contrast removing the scattering effects that had previously shrouded fine features. A variation to remove this scattered light used the HiLo method, also with intensity encoding [76] to increase the signal to noise in the focal region. All of these methods demonstrated successful results but do not seem to have been taken up as standard improvements by the majority of the SPIM community.

Having pondered on the use of intensity modulation on the beam the other consideration was to increase the length over which the light sheet is narrow in effect to produce a larger field of view for each optical slice. This was achieved using a Bessel beam with scanning [77] which has an extended depth of focus but with the cost of redistributing some of the light intensity from the sheet to side lobes, which excite some fluorescence in either side of the main focal plane. Yet, this complication was then removed through the use of the same beam profile, but exploiting a near infrared ultra-short pulse excitation beam to produce two photon excitation [78]. The use of non-linear methods meant that, when the Bessel beam profile is squared (the intensity profile that in effect excites the sample) the extra energy in the side lobes is reduced, compared to the main excitation central core of the beam. In this later setup, the team also added structure illumination to the beam to further

remove the out of focus effects and improved the axial resolution to sub-micron dimensions for the first time. The samples used in this case were cultured cells and not intact whole animals and thus the macroscopic imaging capability of the SPIM system were not utilised, only the speed for optical sectioning and the reduced levels of bleaching. A subsequent paper described in detail how to implement this method in some detail [79], though the optical configuration is still complex and although can clearly be made to work, maintaining this in a standard biological imaging facility would be challenging. This technique was then taken one stage further by applying a 2 dimensional lattice light sheet to enable super-resolution imaging [80]. Again, the results were predominantly from cell cultures or *C. elegans* at a very early (2 cell) stage of development. In this case, the high resolution and still reasonable speed enabled protein movements within the cell to be studied for the first time, and provided some stunning new images.

Building on the use of the a more exotic beam profile, an Airy beam was then introduced [81], offering a simpler implementation into an OpenSpim instrument [82]. The challenge with such a beam format is that it has a curved beam profile with a number of areas of peak intensity, however, it also has the self-healing property of the Bessel beam and extended depth profile. Rather than use non-linear excitation to remove the imaging artefacts introduced by these intensity lobes, deconvolution was applied to the images. This did clean up the images to provide some impressive datasets, but as the full advantages of such a format have yet to be realised. Their true application may well be imaging very deeply within samples when the method may need to be recombined with adaptive optics on the imaging arm of the microscope to remove the aberrations induced on the way out. The deconvolution method they use also assumes that the lobes are in exactly the position predicted, which could lead to spurious features on an image in some situations though this has not yet been shown.

Beyond the use of novel beam profiles, a number of other variations are associated with the sample positioning. The original SPIM configuration used a vertically mounted sample which works well for extended imaging of certain samples. However, this is not suitable for high throughput methods, nor for the use of light sheets in more conventional samples (as mounted for traditional microscopy methods). Thus, results have been reported on inverted and inclined SPIM setups, and dual direction inverted SPIM, as well as their variations, incorporating Raman, two-photon and fluorescence lifetime imaging. Novel methods of introducing the light sheet have also been developed, including the use of a mirror so that the illumination passes through the same objective, or just below it to produce the light sheet [83,84] (a method employed by Leica as an add-on to a confocal to undertake light sheet microscopy).

In order to present the reader with a useful guide to all of these methods they are summarised in Table 1.

Name of the technique	PRINCIPAL FEATURE	Ref.
CLASSIC TECHNOLOGIES		
Ultramicroscopy	First light sheet microscope, used a convergent beam of intense sunlight, and projected a slit through a colloidal sample.	[3]
OPFOS	Orthogonal-plane fluorescence optical sectioning microscopy. Early version the light sheet fluorescent microscope, used a simple cylindrical lens.	[4]
Confocal theta microscopy	Early version, used two objectives to separate the illumination and detection, the observed volume was spherical.	[5,6]
SPIM	Single or Selective Plane Illumination Microscope basic setup, put the technique at the scientific spotlight.	[1]
OPTiSPIM	Integrating optical projection tomography in light sheet microscopy, by rotating the sample to collect 3D data.	[27]
SCANNED BEAM STRUCTURE ILLUMINATION TECHNIQUES		
DSLIM	Digital Scanned Light-Sheet Microscopy, fast scan a laser beam to create a virtual light sheet.	[31]
SPIM with SI	SPIM with structured light	[85]
DSLIM-SI	DSLIM with structured illumination	[75]
3D HiLo SPIM	Additional structuring of the illumination (High and Low frequencies) for improved background rejection	[76]
sTSLIM	Scanning thin-sheet laser imaging microscopy with SI and HiLo background rejection	[86]
Lattice sheet SPIM	Ultrathin light sheets derived from 2D optical lattices, using an SLM to create the lattice and dithered mode to excite the sample, and requires at least three raw images per reconstructed image.	[80]
EXOTIC BEAMS TECHNIQUES		
Bessel SPIM	Scanned Bessel beam plane illumination microscopy using different approaches	[79,80,87–91]
Airy SPIM	Scanned Airy beam plane illumination microscopy using different approaches	[81,82,92,93]
CONFOCAL AND MULTIVIEW TECHNIQUES		
Confocal SPIM	Confocal line detection, single or multiview	[34,35]
diSPIM	Dual view inverted SPIM imaging, requires two raw images per reconstructed image, and deconvolution for a maximum resolution.	[94,95]
HR-OPFOS	High Resolution OPFOS, by scanning an object through the focal zone a and recording only a few 1-D pixel lines within this zone	[96]
mSPIM	Multidirectional SPIM, illuminates sample from both sides	[30]
MuVi-SPIM	MultiView SPIM, two detection and illumination objective lenses	[36]
IsoView SPIM	Each arm can be used for illumination and detection sequentially or simultaneously, giving 4 different views of the sample	[74]
SiMView SPIM	Simultaneous multiview, it is a MuVi-SPIM were both side are simultaneously registered	[73]
triSPIM	Light sheet microscopy with isotropic super-resolution as 3 lenses are disposed in the 3-axis around the sample.	[97]
ASLM	Axially swept light sheet microscopy, using a CMOS camera and a high NA remote focusing objective, scanning the sheet across the sample and registering just the focus region	[98]
DiaSLM	ASLM capable of translating a light-sheet arbitrarily in 3D	[99]
INVERTED, INCLINED AND SINGLE OBJECTIVE TECHNIQUES		
π SPIM or π SPIM	Oblique light-sheet microscope with high NA, high resolution isotropic light-sheet imaging	[100]
Open-top SPIM	SPIM for conventional mounted specimens, lenses at the bottom, imaging at 45 degrees through a water prim and cover glass	[101]
iSPIM	Inverted SPIM for direct imaging of a variety of samples. The lenses are positioned on top of the sample, as in a direct microscope, at 45 degrees.	[102,103]
iSPIM-FCS	inverted SPIM with Fluorescence Correlation Spectroscopy (FCS)	[104]
HILO or iSPIM	Highly Inclined and laminated Optical sheet or inclined SPIM	[105,106]
LSBM	Light-sheet Bayesian super-resolution microscopy, SPIM for super-resolution imaging using a prism-coupled condenser design	[107]
OPM	Optically sectioned imaging by oblique plane microscopy	[108]
RLSM	Reflected light-sheet microscopy, using a micro prism or mirror to reflect	[109,110]

	the light sheet	
SoSPIM	Single Objective SPIM, uses a single objective with a miniature mirror inserted into the objective to redirect the light sheet.	[83]
APOM	Axial Plane Optical Microscopy, use only one objective to create the light-sheet and collect the fluorescence by optically rotating the imaging plane.	[111]
SCAPE	Swept confocally-aligned planar excitation microscopy, use only one objective to create the light-sheet and collect the fluorescence by optically rotating both the imaging and illumination plane.	[112]
ABERRATION CORRECTION TECHNIQUES		
WAO-SPIM or Adaptive SPIM	Imaging deep and clear in thick inhomogeneous samples with a wavefront sensor adaptive optics in the imaging arm of the SPIM	[37–41,113,114]
SPED- SPIM	Spherical-aberration-assisted Extended Depth-of-field light sheet microscopy	[115]
NON-LINEAR, SUPER RESOLUTION AND ADVANCED TECHNIQUES		
2P- SPIM	Combines two-photon imaging with the light-sheet microscope, as well as other technologies to increase resolution and quality	[90,116–126]
Raman SPIM	SPIM integrated with Raman imaging	[127–129]
SPIM-FCS	Fluorescent correlation spectroscopy integrated into a SPIM system using a high speed EMCCD.	[130,131]
SPIM-FLIM	SPIM with Fluorescence Lifetime Imaging Microscopy (FLIM)	[132]
Hyperspectral SPIM	Scanned line SPIM with the detection of the line on a spectrometer	[133]
SPIDDM	Selective-plane illumination differential dynamic microscopy, SPIM combined the digital Fourier methods to provide the ensemble transport dynamics.	[134]
SPIM with trapping	Integrate LSFM which solely uses optical forces to trap and hold the sample using a counter-propagating laser beam geometry.	[135]
IML-SPIM	Individual Molecule Localization SPIM, allowing live 3D super resolution microscopy.	[136–138]
SMD-SPIM	SPIM for Single Molecule Detection and tracking, using single molecule algorithms to analyse the images	[139–142]
SPIM-STED	SPIM with Stimulated Emission Depletion.	[143–146]
RESOLFT	Implements the Reversible Saturable/Switchable Optical (fluorescence) Transitions technique in a light sheet configuration for super-resolution imaging, with axial resolution enhancement.	[147]

In references [8,51,148,149], some of these different SPIM configurations are reviewed, including the exotic beams and two-photon methods. In reference [20], the advantages and disadvantages of some of these techniques are discussed, and in references [84,150] there are further insights on super-resolution and advanced LSFM techniques. For more information on the advanced and single-objective techniques, one can refer to [151], which review LSFM for multiscale imaging.

5. Dynamic SPIM for extended time or rapid viewing of biological events

As illustrated above, one of the strengths of light sheet microscopy lies in both the speed and duration with which one can image and it is therefore the method of choice for non-invasive, temporal imaging of intact, fully functional organisms to study the dynamics involved in shaping complex organisms. In an example study, using a DSLM, the dynamics of *Arabidopsis thaliana* samples was investigated [152], in close to natural growth conditions, allowing imaging live plants at the structural, cellular and subcellular scales. The plant's growth was imaged for multiple days with no perceived damage or alteration to the sample, allowing the study of its cell differentiation and root development. In the case of plants, which clearly use light as part of their development processes, the excitation light for SPIM was only activate as an optical slice was obtained, thus minimising the effects of the imaging light on the growth of the plant. SPIM offers the best compromise between preserving conditions as close as possible to those natural to the sample, and allowing the

observation of biological processes at the appropriate temporal and spatial resolution [12,14].

This extended time imaging of SPIM, therefore, utilises the minimal light dose to the sample among the light sheet methods, however this leads to a new challenge, to maintain the sample under physiological conditions. In the case of Zebrafish imaging, mSPIM was used for long-term imaging [153] where growth and immobility defined the criteria for proper development, and spatial confinement, for precise imaging of the zebrafish embryo. The team used 3% methylcellulose or 0.1% agarose in an FEP tube to ensure proper growth and immobilization. The other secret being to maximise the light collection in order to reduce the excitation intensity.

In analogy to using the extended imaging capability of SPIM, is to use its speed to capture an optical slice. This speed of image capture has been used in different ways to watch high-speed dynamic processes, as for example, to monitor the movement of the beating heart in the zebrafish. In one approach, a conventional near infrared transmission image is taken on a camera at around 100 frames per second and analysed in real-time for the movement pattern of the heart. After a few cycles the system is capable of predicting when the heart will be in a certain position in its cycle, and this predication is used to activate the light sheet and camera to record an optical slice at a particular point in the heart cycle [154]. Based upon this, a dynamic image of the beating heart can then be recorded of movement in three dimensions. An example of the images such method can obtain is showed in figure 4, presenting the zebrafish heart in different configurations. An alternative approach is to record multiple images at high speed and then to use post processing to assemble the movement by looking for similarities within images at different time points [155]. In this latter publication, a new insight into the role of certain proteins in the sheer stress within the heart was provided.

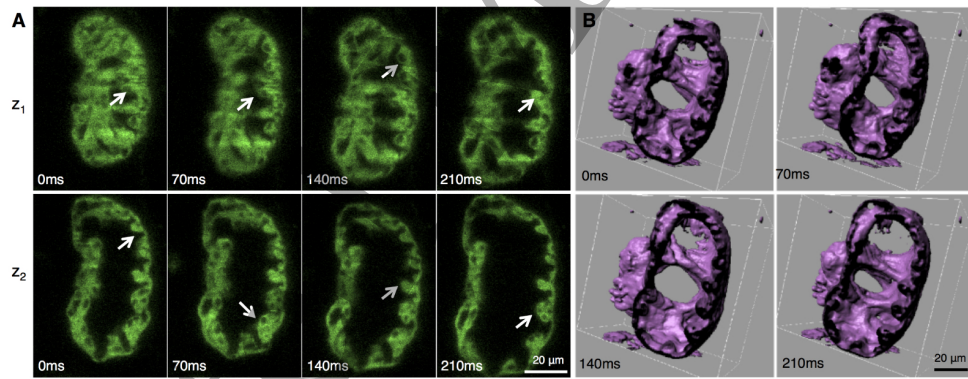


Figure 4. Dynamic image of the beating heart. LSFM images of the zebrafish heart, at different positions in the beating cycle. (a) Fluorescent images of the heart at different point in the beating cycle, and (b) 3D rendering of the zebrafish heart at different points in the beating cycle. This image is from Trivedi et al 2015 [159].

The other area of high-speed imaging is to create rapid three-dimensional data sets. Using a conventional SPIM system, though with dual excitation and collection, it has been possible to image at a rate of 200 frames per second [78]. This lead to a 50-slice volume taking around 0.5s to record. However, this turned to a significant challenge, when imaging the neural wiring during *C. elegans* development for five-hour period [95], of vast quantities of data. Crucial to the biological relevance in this was the low phototoxicity of the SPIM method. At present, the highest recorded speed for volumetric imaging is through a variation of the SPIM method, in a system known as SCAPE [112]. Figure 5 illustrates the key optical

concepts in the SCAPE method which achieved imaging at rates up to 200 volumes per second and been used to study signalling within the mouse brain. This method uses a single objective and the light sheet is scanned through the lens at an angle using a polygon mirror. The objective collects the fluorescence light, which is then reflected onto the camera by an adjacent element on the polygon array. This has the effect of sweeping the light sheet rapidly through the volume of the sample with only the rotation of the mirror. The method was also demonstrated in the same publication in *Drosophila*, to observe the beating heart.

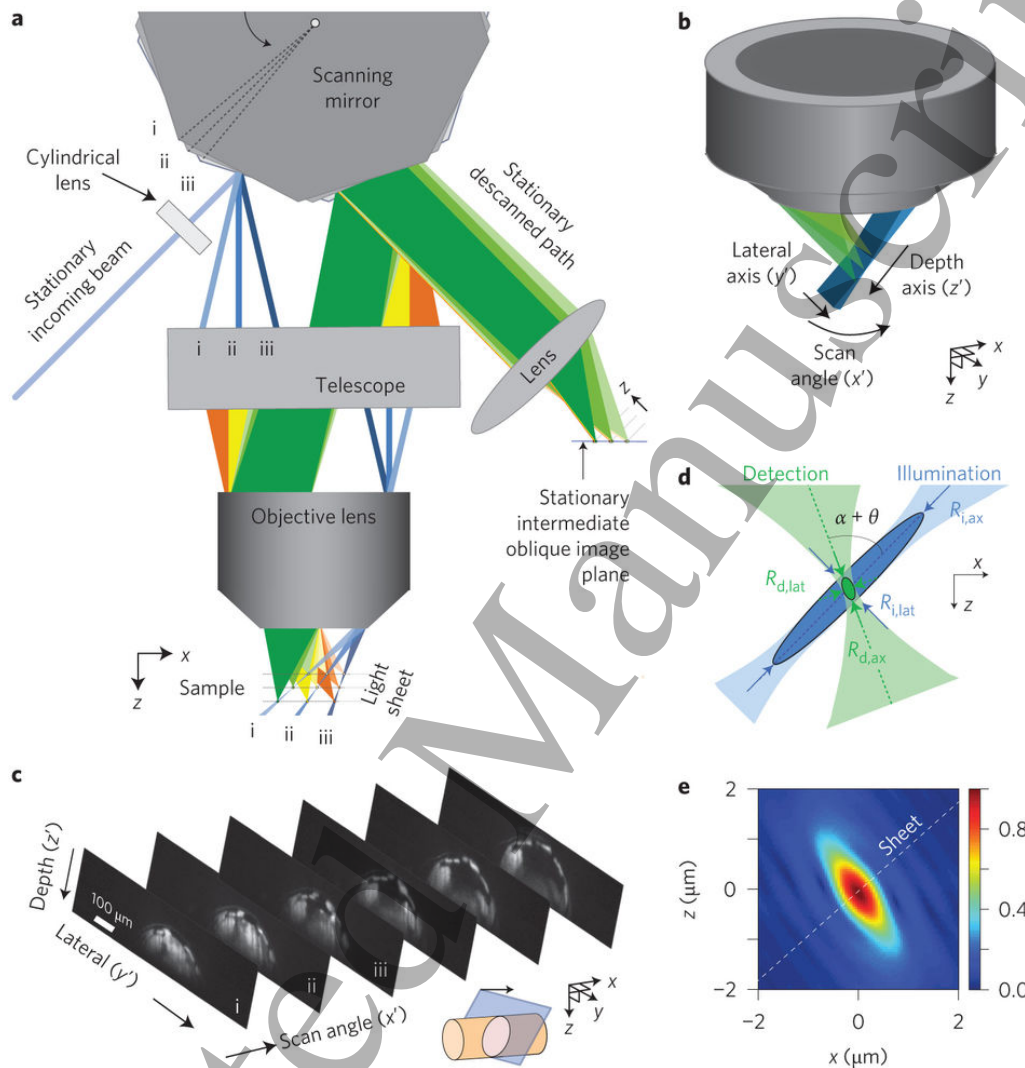


Figure 5. Swept confocally-aligned planar excitation (SCAPE) microscopy from Bouchard et al 2015 [115].

6. Advanced SPIM: non-linear and super-resolution techniques

As mentioned before, the main advantages of LSFM are related to its optical sectioning, high contrast, high imaging speed, and low phototoxicity [17], but every technology has its drawbacks [149,156]. Because of its inherent flexibility, light sheet microscopy can be adapted to overcome specific challenges in many different configurations, as it is a mesoscopic method that uses an engineered light sheet, of finite thickness, to excite fluorophores within its volume. Although, it is the least invasive, in terms of excitation light intensity, compared to other routine microscopy methods, and allows near-physiological monitoring of intracellular dynamics at very fast rates and for a prolonged imaging period, its

resolution is not as high as some methods, in effect, trading speed for resolution. But LSFM is such a versatile technology that it can be combined with other techniques to overcome these resolutions limitations, such as two-photon imaging, structured illumination or STED to reach super-resolution levels. These methods have then been linked with other detection methods, such as Raman, to obtain multispectral images [157] at high resolution, though generally at the expensive of temporal resolution.

One of the first moves for improved resolution, and an increase in the potential depth at which samples can be imaged, was the use of non-linear excitation. In conventional beam scanned non-linear microscopy this typically leads to a loss of resolution compared to single photon excitation due to the longer wavelength increasing diffraction effects. However, in the case of SPIM the non-linear effect can be used as an advantage, reducing the volume in which the light intensity is high enough to stimulate excitation. Development of the light-sheet excited multiphoton fluorescence microscopy has now been reported by several research groups [90,116–126,137]. These instruments frequently combine other techniques to increase resolution and quality, such as incorporating Bessel beams and structured illumination as well as using Second Harmonic generated images. Other technologies that have already be integrated into SPIM are Raman imaging [127–129], Fluorescent correlation spectroscopy [130], Fluorescence Lifetime Imaging Microscopy (FLIM) [132], spectral imaging [133]. These multimodal systems represent an optimal platform for tissue imaging and they are further increasing the applicability of light sheet microscopy to a wider range of samples, including those that have to be mounted on non-transparent surfaces.

The next obvious direction for LSFM was to follow the path into super-resolution imaging, yet clearly recognising, again, that the potential for high-speed imaging was being traded-off against resolution. The first advance here was through the various methods of imposing a light intensity pattern on the excitation beam which was subsequently followed by a more direct implementation of the structured illumination method [85,75]. Although the method led to an improvement in resolution, the main benefit was a reduction in signal from scattered light, leading to enhanced contrast images. This work was then followed by true super-resolution imaging in a LSFM using the PALM technique [136,138]. In this work two light beams were used, one to activate a reduced set of the fluorescent molecules within the sample and subsequently to de-activate and then activate a separate set. To this date, this method has only been used in tumour spheroids to a depth of around 150 microns and not in more intact samples. The Stochastic super-resolution method, STORM, was combined with SPIM in 2015 to a yeast based sample [139]. Further variations on these methods, all using localisation algorithms to determine a single molecule's position have since been implemented [136–142] in a broader range of samples.

Light sheet implementation of STED was achieved in 2011, followed again by variations in the method [143–146]. The original version is worthy of specific mention, as they used a dye that emitted in the near infrared, as scattering was a potential confounding issue and the longer wavelength helped to reduce this effect. The aim was to increase the resolution along the imaging optical axis, and a light sheet was created with a laser around 640 nm and this sheet was surrounded by a sheet shaped for depletion beam, with a hole through the middle (in effect very elongated ellipse with a central hole) created using a TEM_{01} beam profile sent through the cylindrical lens in a conventional SPIM system. This gave a 60% improvement in their axial resolution. In another variation of the method, and perhaps closer to the PALM technique, RESOLFT (reversible saturable/switchable optical fluorescence transitions) has been implemented in a light sheet configuration [147]. Here, rather than causing stimulated emission to remove the excited molecules, photoactivatable molecules are switched off such that only a thin sheet of molecules is available for excitation by the light sheet. Once sufficient photons have been gathered for one very thin section the next layer can be prepared with, again, only a thin sheet of molecules being allowed to enter the fluorescent emission state.

In an alternative method, rather than removing the emission or preventing the excitation of molecules, the volume in which they were created was minimised in the so called Lattice Sheet microscope [80]. The full details on how this was achieved can be found in the publication, but in simple terms, through the interferences of light patterns, it is possible to create a light sheet consisting of multiple point in space and by dithering these spots around, it is possible to fill an optical plane to take a slice where each excited pixel is very small. Using this method, it was possible to image at high frame rates though some level of post processing is required. To date, this is the highest speed, super-resolution method that has been developed, however full implementation is not trivial, both optically and computationally. A review of the super-resolution methods and specifically their application to plants can be found in [158].

7. Some Selection Considerations

As is clear from the text above, that the number of light sheet microscope options is now enormous, with each method having its own specific application, advantages and drawbacks. In this section, we will try and provide some guidance to the reader on the considerations that should be made before selecting a particular method. As with all imaging tasks, the final selection is certain to involve some form of compromise between competing parameters, such as resolution, speed, field of view, ease-of-use and perhaps, in the end, cost.

Before discussing the selection of a specific light sheet method, it is worth recapping briefly on why you might consider using this approach in preference to other three-dimensional imaging methods. One clearly needs a sample that is generally transparent, not too highly scattering and has three-dimensional structure. The major advantages of the light sheet method are the ability to record optical slices rapidly with high, but generally not super, resolution. The images are normally recorded with lower levels of phototoxicity and photobleaching than alternative three-dimensional methods and this makes extended imaging of live samples more viable. With the advent of the OpenSPIM options it is also the lowest cost three-dimensional imaging modality.

The first consideration is the main configuration of the light sheet that one wishes to use and this is entirely determined by the sample. The conventional light sheet, with the optics in the plane of the table, requires the sample to be suspended in centre of the instrument in a clear material. Although this is the most common light sheet configuration, the challenges in mounting the sample should not be overlooked. For Zebrafish, plants and organoids mounting inside agar or FEP tubing works well and provides access to all of the major areas within the sample. *C. elegans* can also be imaged in this manner but are perhaps more amenable to horizontal sample mounting. Referring to table 1, this is the case of configurations based on the classic technologies configuration, as is the case of SPIM and DSLM. Optically the conventional configuration is the easiest system to build but is not the best system for high-throughput imaging, as each sample has to be individually prepared. If the sample is not highly fluorescent then adding a second imaging camera and light beam to increase the light intensity is most easily achieved in the conventional configuration. The second light sheet is also useful for imaging more deeply in samples, or samples where excitation light can be lost before reaching the target organs of interest.

The inverted and inclined configurations require greater engineering skills to bring the optics out of the plane of the table, as they are often perpendicular. This configuration does enable the sample to be laid down in the focal volume and thus the sample could be in microfluidic channels and moved more rapidly under the imaging instrument. Most of the inverted, incline or single objective techniques allow the sample to be placed on top of microscope slides, as is the case of iSPIM, open-top SPIM and SCAPE referred in table 1.

The inverted configuration is perhaps better for brain imaging in a range of samples, including rodents, and thus if this is the main imaging target, the extra complication of the inverted method is worth considering.

Having decided on the configuration, now one has to consider the resolution and field of view required. Many systems, including the ones built by the authors, use a x10, 0.3 NA excitation objective and a x16, 0.8 NA imaging objective. This, in theory, provides a field of view of around 1.5 mm. However, it is not possible to produce a uniform, thin light sheet over this volume, unless a Bessel beam is utilised (with the other complications of side lobes which need to be removed, normally by post processing) or other advanced techniques as ASLM. For a light sheet of 2 microns thickness one typically has a field of view of around 150 microns. When the thickness of the light sheet is convolved with the point spread function of the imaging objective this gives an axial resolution of around 1.5 microns before any post processing is undertaken. For high axial resolution one can make the light sheet thinner but the region over which it is thin and uniform decreases rapidly.

The highest lateral resolution that one can achieve is limited by the imaging objective NA and is around 0.4 to 0.5 microns. To achieve this performance one requires a camera with sufficient pixels of the correct size for Nyquist sampling of the field of view. For example, for 0.5 micron resolution and 16 times magnification, one would require pixels of around 4 microns for Nyquist sampling. These criteria are now easily reached using modern sCMOS cameras. Again, one can increase the field of view, by adjusting the tube lens before the camera, which alters the effective magnification but will then no longer be Nyquist sampled and this can introduce distortions in the image.

The next decision is likely to be on the speed and volume that one wishes to image. Higher speed is probably best achieved at present, either using the SCAPE configuration, or the use of two cameras, one from each direction, and two light beams synchronised so that while one camera is imaging, the other is transferring its previous frame, as in MuVi-SPIM. It should be remembered that in high speed imaging, one does need a bright sample and that the signal to noise will become worse, but this is the expense that has to be paid for the speed. It is likely that the volume imaged will decrease, again so that the region imaged is seen multiple times per second. When imaging at high speed, it is also vital that the data handling is considered at the outset, both in terms of storing the data and also subsequent processing.

In the foregoing text we have also described multiple other features that have been developed to overcome imaging challenges. Should these be employed on a routine basis? The use of the scanned light sheet is useful in samples where there are absorbing features present before the area of interest. This method does remove “streaks” in the image and should probably be considered for most samples. Dual direction illumination also helps with this issue, though adds significantly to the complexity of alignment. In samples that are more highly scattering, and when imaging more deeply, then the use of the scanned light beam combined with the rolling shutter to produce effectively a line scanned confocal system is a very powerful option, such in confocal SPIM. Again, this requires greater instrumentation expertise but the removal of scattered light from the image can significantly improve the signal to noise. The exact gain though is clearly sample dependent.

The final areas, and the point at which the cost of systems significantly increase, is in the use of non-linear excitation and super-resolution. Both are more technically challenging and are probably only required for specific imaging applications. Non-linear excitation does enable deeper imaging, but unlike beam scanned non-linear microscopy, where all fluorescent photons can be collected as they can only have come from a single volume in the sample, scattering of the fluorescent light as it emerges through the sample limits non-linear methods usefulness. The combination of the rolling shutter and non-linear excitation

can help to mitigate scattering on the output but one is now rejecting precious signal photons. Super-resolution, using either structured illumination or STORM/PALM options, should only be considered when the extra resolution is really required to understand the biology taking place. One generally loses speed and the low photobleaching advantages of SPIM and thus perhaps the advantages are out-weighed by these losses.

8. Future Directions and Summary

As can be seen from the previous sections, light sheet microscopy has made huge advances in the last few years, since its original inception, and this has been strongly influenced by advances in technology, both optically and biologically. The question then has to be asked: what might come next? Presently, within the field of Biophotonics, the term “optogenetics” is being used widely, and the main application area has been in the field of neuroscience. Where by genetically encoding a protein, which can be “switched on” or “off”, the reactions of a live animal can be altered by using light. This means that specific neural pathways can be activated and studied in isolation and light sheet microscopy, as mentioned in numerous places earlier, is now becoming one of the methods for studying brain activity with its ability for high-speed 3D imaging [112]. Another variation on this is the use of light to specifically kill cells that are expressing the “KillerRed” protein [159]. Here, when exposed to light around 561nm the cells expressing the protein die due to the production of reactive oxygen species during the fluorescence process. In previous works, this transfection and subsequent cell death was only undertaken in cell based studies. However, using the SPIM configuration, with the “KillerRed” being expressed in the kidney’s renin producing cells, live zebrafish were treated with a high survival rate, but presenting changes in physiology due to the loss of these cells [160]. The team used both the SPIM sheet to excite a larger number of cells, or a Bessel beam introduced through the imaging arm. Use of conventional optical microscopes led to high death rates in the fish. This was not the first time that a SPIM configuration had been used for what might be described as “three-dimensional laser microsurgery” [71], where direct ablation of cells was achieved or local photo-activation of cells was undertaken [66].

A further area of direct biological application of SPIM is towards the use of three-dimensional cell clusters or, more especially, organs extracted from deep within an animal. Within the SPIM system it has been shown that it is possible to keep entire organs alive for extended periods of time, and to use the light sheet to study cell interactions. This is likely to have a specific role to play in the field of immunology, where the organs are frequently hard to access using conventional methods, but where the intact organ continues to function for some time outside the animal [161]. Thus, as well as the continued imaging of entire samples, such as Zebrafish and Drosophila, the imaging of intact, normally deep seated organs is likely to be an area of growing interest. The other area being explored by several groups is the use of SPIM, typically the inverted configuration, for high throughput screening and commercial systems are looking to use a micro-fluidic system to direct the samples, typically Zebrafish, through the SPIM imaging volume [162,163], or to perform cell cytometry [164].

In terms of the optical developments in light sheet based imaging, the drive is likely to continue to be in two directions. The first is for increased speed, and the SCAPE system, described above, is currently the leading implementation of this method. There will eventually be a limit on the speed as insufficient photons will be available for high quality imaging, but at the moment, the bottleneck is in the transfer of the images from the camera, although frame-rates are rising rapidly. The other area is likely to be improved resolution and contrast. As described, a wide variety of beam configurations have been used, including complex lattices of light. These have improved both the imaging contrast and spatial resolution, but typically at the expense of imaging speed. As the move is towards ever

deeper imaging, in more mature samples, the further application of adaptive optics is likely to be important. Their use on the imaging arm has already been described [40], and it has been shown that they can be used to optimise an excitation beam profile [113], spatial adjustment of the excitation beam to directly overcome sample induced aberrations in the excitation beam path has only been recently reported [114]. An alternative approach is also to build the light sheet system on the end of an optical fibre, for *in vivo* invasive light-sheet microscopy at the tip of a needle [165]. It is likely that this regime of in depth imaging will be an area of technical advance in the near future. As more life-scientists really appreciate what light sheet imaging is capable of, then, it will be applied to an ever-increasing range of samples which are likely to require some novel optical solution to achieve the results required to answer increasing complex 4D challenges.

Computers and data processing is also playing a growing role in the field, as all imaging methods and protocols are already being developed to improve the apparent resolution and contrast within images [166]. However, one area that is only just being considered, however all builders and users of SPIM instruments were well aware of, is the challenge presented by the very large datasets that can be generated. Although all scientists are currently driven to store, and subsequently make available, their raw data before any manipulation or analysis is undertaken, this situation may need to change. With single data sets for a time lapsed imaging session, it is easy to take 3 to 4 Tb, therefore the storage requirements are becoming excessive and perhaps only processed data sets may be stored in the future. The different computational steps used in all imaging are now being quantified and examined in detail, and the inherent challenges and the overall status of available software for bioimage informatics, focusing on open-source options is now an important topic [167] and protocols and software for data management, processing and analysis [168] and data processing [169] are being discussed. To achieve the maximum benefit from this work, the data processing aspects must be considered alongside the optical advances in a truly integrated manner.

Since its re-invention light sheet microscopy has advanced hugely in terms of the quality, range of samples imaged and speed of image capture. What is now routine would have been totally unbelievable to the original inventors over a hundred years ago. The light sheet and associated optics is the latest method, revolutionising the understanding of complex biological processes, crucially bringing both high resolution, three-dimensional imaging and speed, so that dynamic processes can be studied and understood in detail. This has been brought about over the integrated work between optical physicists, instrument builders, commercial technology suppliers, and the end users and the Open-SPIM model of releasing full designs of working systems that can be built at low cost from commercial components is a very interesting model. Indeed the Open-SPIM route may become more common in the future as a result of its success and also as it becomes easier to purchase and combine high performance optics and computer controlled cameras and positioning equipment. Light sheet microscopy is now really starting to answer complex questions that previously could not even be considered, including how we all think and live.

Acknowledgements

The authors would like to acknowledge the support of multiple colleagues who have helped them within the field of light sheet microscopy, specifically Dr Laura Young, Dr Chris Saunter, Dr Miguel Jarrin and Prof Roy Quinlan in Durham University, Dr Charlotte Buckley, Dr Sebastian Rider and Prof John Mullins at Edinburgh University and Dr Jonny Taylor now at Glasgow University. Funding supporting this work has come from the Engineering and Physical Sciences Research Council in the UK, British Heart Foundation, Wellcome Trust, Leverhulme Trust and the Brazilian agencies, CNPq and CAPES.

References

- [1] Huisken J, Swoger J, Del Bene F, Wittbrodt J and Stelzer E H K 2004 Optical Sectioning Deep Inside Live Embryos by Selective Plane Illumination Microscopy *Science* (80-.). **305** 1007–9
- [2] Siedentopf H and Zsigmondy R 1902 Über Sichtbarmachung und Größenbestimmung ultramikroskopischer Teilchen, mit besonderer Anwendung auf Goldrubingläser *Ann. Phys.* **315** 1–39
- [3] Mappes T, Jahr N, Csaki A, Vogler N, Popp J and Fritzsche W 2012 The Invention of Immersion Ultramicroscopy in 1912-The Birth of Nanotechnology? *Angew. Chemie Int. Ed.* **51** 11208–12
- [4] VOIE A H, Burns D H and Spelman F A 1993 Orthogonal-plane fluorescence optical sectioning: Three-dimensional imaging of macroscopic biological specimens *J. Microsc.* **170** 229–36
- [5] Stelzer E H K and Lindek S 1994 Fundamental reduction of the observation volume in far-field light microscopy by detection orthogonal to the illumination axis: confocal theta microscopy *Opt. Commun.* **111** 536–47
- [6] Stelzer E H K, Lindek S, Albrecht S, Pick R, Ritter G, Salmon N J and Stricker R 1995 A new tool for the observation of embryos and other large specimens: confocal theta fluorescence microscopy *J. Microsc.* **179** 1–10
- [7] Fuchs E, Jaffe J, Long R and Azam F 2002 Thin laser light sheet microscope for microbial oceanography. *Opt. Express* **10** 145
- [8] Santi P A 2011 Light Sheet Fluorescence Microscopy *J. Histochem. Cytochem.* **59** 129–38
- [9] Buytaert J A N, Descamps E, Adriaens D and Dirckx J J J 2012 The OPFOS Microscopy Family: High-Resolution Optical Sectioning of Biomedical Specimens *Anat. Res. Int.* **2012** 1–9
- [10] Shimomura O, Chalfie M, Kenan W R and Tsien R Y 2008 The Nobel Prize in Chemistry 2008 'for the discovery and development of the green fluorescent protein, GFP' *Nobelprize.org*
- [11] White R M, Sessa A, Burke C, Bowman T, LeBlanc J, Ceol C, Bourque C, Dovey M, Goessling W, Burns C E and Zon L I 2008 Transparent Adult Zebrafish as a Tool for In Vivo Transplantation Analysis *Cell Stem Cell* **2** 183–9
- [12] Berthet B and Maizel A 2016 Light sheet microscopy and live imaging of plants *J. Microsc.* **263** 158–64
- [13] Ovečka M, Vaškebová L, Komis G, Luptovčiak I, Smertenko A and Šamaj J 2015 Preparation of plants for developmental and cellular imaging by light-sheet microscopy *Nat. Protoc.* **10** 1234–47
- [14] Stelzer E H K 2014 Light-sheet fluorescence microscopy for quantitative biology *Nat. Methods* **12** 23–6
- [15] Pawley J B 2006 *Handbook of biological confocal microscopy*
- [16] Girkin J M and McConnell G 2005 Advances in laser sources for confocal and multiphoton microscopy *Microsc. Res. Tech.* **67** 8–14
- [17] Stelzer E H K 2009 Light sheet based fluorescence microscopes (LSFM, SPIM, DSLM) reduce phototoxic effects by several orders of magnitude *Mech. Dev.* **126** S36–S36
- [18] Park O K, Kwak J, Jung Y J, Kim Y H, Hong H-S S, Hwang B J, Kwon S-H H and Kee Y 2015 3D Light-Sheet Fluorescence Microscopy of Cranial Neurons and Vasculature during Zebrafish Embryogenesis *Mol. Cells* **38** 975–81
- [19] Keller P J, Pampaloni F and Stelzer E H K 2006 Life sciences require the third dimension *Curr. Opin. Cell Biol.* **18** 117–24
- [20] Follain G, Mercier L, Osmani N, Harlepp S and Goetz J G 2017 Seeing is believing – multi-scale spatio-temporal imaging towards in vivo cell biology *J. Cell Sci.* **130** 23–38
- [21] Huisken J 2005 Studying live embryos in 4D: selective plane illumination microscopy: a new technique for high-resolution four-dimensional imaging *Biochem. (Lond).* **27**

- 13–6
- [22] Verveer P J, Swoger J, Pampaloni F, Greger K, Marcello M and Stelzer E H K 2007 High-resolution three-dimensional imaging of large specimens with light sheet-based microscopy *Nat. Methods* **4** 311–3
 - [23] Huisken J, Swoger J, Lindek S and Stelzer E H K 2006 Selective plane illumination microscopy *Handbook of Biological Confocal Microscopy* pp 672–9
 - [24] Greger K, Swoger J and Stelzer E H K 2007 Basic building units and properties of a fluorescence single plane illumination microscope *Rev. Sci. Instrum.* **78** 23705
 - [25] Mohan K and Mondal P P 2014 Experimental Verification of Vectorial Theory to Determine Field at the Geometrical Focus of a Cylindrical Lens *Microsc. Res. Tech.* **77** 105–9
 - [26] Reynaud E G, Krzic U, Greger K, Stelzer E H K, Kržič U, Greger K and Stelzer E H K 2008 Light sheet-based fluorescence microscopy: More dimensions, more photons, and less photodamage *HFSP J.* **2** 266–75
 - [27] Mayer J, Robert-Moreno A, Danuser R, Stein J V., Sharpe J and Swoger J 2014 OPTiSPIM: integrating optical projection tomography in light sheet microscopy extends specimen characterization to nonfluorescent contrasts *Opt. Lett.* **39** 1053
 - [28] Abdellah M, Bilgili A, Eilemann S, Markram H and Schürmann F 2015 Physically-based in silico light sheet microscopy for visualizing fluorescent brain models *BMC Bioinformatics* **16** S8
 - [29] Keller P J and Dodt H-U 2012 Light sheet microscopy of living or cleared specimens *Curr. Opin. Neurobiol.* **22** 138–43
 - [30] Huisken J and Stainier D Y R 2007 Even fluorescence excitation by multidirectional selective plane illumination microscopy (mSPIM) *Opt. Lett.* **32** 2608
 - [31] Keller P J, Schmidt A D, Wittbrodt J and Stelzer E H K K 2008 Reconstruction of zebrafish early embryonic development by scanned light sheet microscopy. *Science* **322** 1065–9
 - [32] Mertz J and Kim J 2010 Scanning light-sheet microscopy in the whole mouse brain with HiLo background rejection *J. Biomed. Opt.* **15** 16027
 - [33] Joubert J and Sharma D 2011 Light microscopy digital imaging *Curr. Protoc. Cytom.* **Chapter 2** Unit 2.3
 - [34] Baumgart E and Kubitscheck U 2012 Scanned light sheet microscopy with confocal slit detection *Opt. Express* **20** 21805–14
 - [35] de Medeiros G, Norlin N, Gunther S, Albert M, Panavaite L, Fiuza U-M, Peri F, Hiiragi T, Krzic U, Hufnagel L, Medeiros G de, Norlin N, Gunther S, Albert M, Panavaite L, Fiuza U-M, Peri F, Hiiragi T, Krzic U and Hufnagel L 2015 Confocal multiview light-sheet microscopy *Nat. Commun.* **6** 8881
 - [36] Krzic U, Gunther S, Saunders T E, Streichan S J and Hufnagel L 2012 Multiview light-sheet microscope for rapid in toto imaging *Nat. Methods* **9** 730–3
 - [37] Girkin J M, Poland S and Wright A J 2009 Adaptive optics for deeper imaging of biological samples *Curr. Opin. Biotechnol.* **20** 106–10
 - [38] Jorand R, Le Corre G, Andilla J, Maandhui A, Frongia C, Lobjois V, Ducommun B and Lorenzo C 2012 Deep and clear optical imaging of thick inhomogeneous samples. *PLoS One* **7** e35795
 - [39] Bourgenot C, Saunter C D, Love G D and Girkin J M 2013 Comparison of closed loop and sensorless adaptive optics in widefield optical microscopy *J. Eur. Opt. Soc.* **8** 13027
 - [40] Bourgenot C, Saunter C D, Taylor J M, Girkin J M and Love G D 2012 3D adaptive optics in a light sheet microscope *Opt. Express* **20** 13252
 - [41] Taylor J M, Saunter C D, Bourgenot C, Girkin J M and Love G D 2015 Realtime wavefront sensing in a SPIM microscope, and active aberration tracking *Proc. SPIE* vol 9335p 93350A–93350A–6
 - [42] Gualda E, Moreno N, Tomancak P and Martins G G 2014 Going ‘open’ with Mesoscopy: a new dimension on multi-view imaging *Protoplasma* **251** 363–72
 - [43] Girstmair J, Zakrzewski A, Lapraz F, Handberg-Thorsager M, Tomancak P, Pitrone P

- G, Simpson F and Telford M J 2016 Light-sheet microscopy for everyone? Experience of building an OpenSPIM to study flatworm development *BMC Dev. Biol.* **16** 22
- [44] Pitrone P G, Schindelin J, Stuyvenberg L, Preibisch S, Weber M, Eliceiri K W, Huiskens J and Tomancak P 2013 OpenSPIM: an open-access light-sheet microscopy platform *Nat. Methods* **10** 598–9
- [45] Gualda E J, Pereira H, Vale T, Estrada M F F F, Brito C and Moreno N 2015 SPIM-fluid: open source light-sheet based platform for high-throughput imaging *Biomed. Opt. Express* **6** 4447–56
- [46] Graham D M 2017 An AutoPilot platform for high-resolution light-sheet microscopy *Lab Anim. (NY)*. **46** 25–25
- [47] Schmied C, Stamatakis E and Tomancak P 2014 Open-source solutions for SPIM image processing *Methods Cell Biol.* **123** 505–29
- [48] Gualda E J, Vale T, Almada P, Feijó J a, Martins G G and Moreno N 2013 OpenSpinMicroscopy: an open-source integrated microscopy platform *Nat. Methods* **10** 599–600
- [49] Preibisch S, Saalfeld S, Schindelin J and Tomancak P 2010 Software for bead-based registration of selective plane illumination microscopy data. *Nat. Methods* **7** 418–9
- [50] Royer L a, Weigert M, Günther U, Maghelli N, Jug F, Sbalzarini I F and Myers E W 2015 ClearVolume: open-source live 3D visualization for light-sheet microscopy *Nat. Methods* **12** 480–1
- [51] Lim J, Lee H K, Yu W and Ahmed S 2014 Light sheet fluorescence microscopy (LSFM): past, present and future *Analyst* **139** 4758–68
- [52] Höckendorf B, Thumberger T and Wittbrodt J 2012 Quantitative Analysis of Embryogenesis: A Perspective for Light Sheet Microscopy *Dev. Cell* **23** 1111–20
- [53] Weber M and Huiskens J 2011 Light sheet microscopy for real-time developmental biology *Curr. Opin. Genet. Dev.* **21** 566–72
- [54] Coelho M, Maghelli N and Tolić-Nørrelykke I M 2013 Single-molecule imaging in vivo: the dancing building blocks of the cell *Integr. Biol.* **5** 748
- [55] Lin H C A, Chekkoury A, Omar M, Schmitt-Manderbach T, Koberstein-Schwarz B, Mappes T, López-Schier H, Razansky D and Ntziachristos V 2015 Selective plane illumination optical and optoacoustic microscopy for postembryonic imaging *Laser Photonics Rev.* **9** L29–34
- [56] Tomer R, Khairy K, Keller P J, Megason S, Srinivas S, Dickinson M, Hadjantonakis A-K, Tomer R, Khairy K and Keller P J 2011 Shedding light on the system: Studying embryonic development with light sheet microscopy *Curr Opin Genet Dev* **21** 558–65
- [57] Khairy K and Keller P J 2011 Reconstructing embryonic development *Genesis* **49** 488–513
- [58] Ahrens M B, Orger M B, Robson D N, Li J M and Keller P J 2013 Whole-brain functional imaging at cellular resolution using light-sheet microscopy *Nat. Methods* **10** 413–20
- [59] Royer L A L A, Lemon W C, Chhetri R K, Wan Y, Coleman M, Myers E W and Keller P J 2016 Adaptive light-sheet microscopy for long-term, high-resolution imaging in living organisms *Nat. Biotechnol.* **34** 1267–78
- [60] Quirin S, Vladimirov N, Yang C-T, Peterka D S, Yuste R, Ahrens M B, B. Ahrens M and Ahrens M B 2016 Calcium imaging of neural circuits with extended depth-of-field light-sheet microscopy *Opt. Lett.* **41** 855
- [61] Costa A, Candeo A, Fieramonti L, Valentini G and Bassi A 2013 Calcium Dynamics in Root Cells of Arabidopsis thaliana Visualized with Selective Plane Illumination Microscopy ed M Bennett *PLoS One* **8** e75646
- [62] Swoger J, Muzzopappa M, López Schier H and Sharpe J 2011 4D retrospective lineage tracing using SPIM for zebrafish organogenesis studies *J. Biophotonics* **4** 122–34
- [63] Mellman K, Huiskens J, Dinsmore C, Hoppe C and Stainier D Y 2012 Fibrillin-2b regulates endocardial morphogenesis in zebrafish *Dev. Biol.* **372** 111–9
- [64] Mickoleit M, Schmid B, Weber M, Fahrbach F O, Hombach S, Reischauer S and

- Huisken J 2014 High-resolution reconstruction of the beating zebrafish heart *Nat. Methods* **11** 919–22
- [65] Jemielita M, Taormina M J, DeLaurier A, Kimmel C B and Parthasarathy R 2013 Comparing phototoxicity during the development of a zebrafish craniofacial bone using confocal and light sheet fluorescence microscopy techniques *J. Biophotonics* **6** 920–8
- [66] Arrenberg A B, Stainier D Y R, Baier H and Huisken J 2010 Optogenetic Control of Cardiac Function *Science (80-.)*. **330** 971–4
- [67] Daetwyler S and Huisken J 2016 Fast Fluorescence Microscopy with Light Sheets *Biol. Bull.* **231** 14–25
- [68] Keller P J 2013 In vivo imaging of zebrafish embryogenesis *Methods* **62** 268–78
- [69] Young L K, Jarrin M, Saunter C D, Quinlan R and Girkin J M 2015 Using SPIM to track the development of the focal power of the zebrafish lens *Optics Express* vol 20, ed A M Rollins, S E Fraser and M A Choma (Zeiss Microscopy GmbH Carl) p 933408
- [70] Lorenzo C, Frongia C, Jorand R, Fehrenbach J, Weiss P, Maandhui A, Gay G, Ducommun B and Lobjois V 2011 Live cell division dynamics monitoring in 3D large spheroid tumor models using light sheet microscopy *Cell Div.* **6** 22
- [71] Engelbrecht C J, Greger K, Reynaud E G, Krzic U, Colombelli J and Stelzer E H K 2007 Three-dimensional laser microsurgery in light-sheet based microscopy (SPIM) *Opt. Express* **15** 6420–30
- [72] Wohland T, Shi X, Sankaran J and Stelzer E H K 2010 Single plane illumination fluorescence correlation spectroscopy (SPIM-FCS) probes inhomogeneous three-dimensional environments. *Opt. Express* **18** 10627–41
- [73] Tomer R, Khairy K, Amat F and Keller P J 2012 Quantitative high-speed imaging of entire developing embryos with simultaneous multiview light-sheet microscopy *Nat. Methods* **9** 755–63
- [74] Chhetri R K, Amat F, Wan Y, Höckendorf B, Lemon W C and Keller P J 2015 Whole-animal functional and developmental imaging with isotropic spatial resolution *Nat. Methods* **12** 1171–8
- [75] Keller P J, Schmidt A D, Santella A, Khairy K, Bao Z, Wittbrodt J and Stelzer E H K 2010 Fast, high-contrast imaging of animal development with scanned light sheet-based structured-illumination microscopy *Nat. Methods* **7** 637–42
- [76] Bhattacharya D, Singh V R, Zhi C, So P T C, Matsudaira P and Barbastathis G 2012 Three dimensional HiLo-based structured illumination for a digital scanned laser sheet microscopy (DSLM) in thick tissue imaging *Opt. Express* **20** 27337–47
- [77] Fahrbach F O and Rohrbach A 2010 A line scanned light-sheet microscope with phase shaped self-reconstructing beams *Opt. Express* **18** 24229
- [78] Planchon T A, Gao L, Milkie D E, Davidson M W, Galbraith J A, Galbraith C G and Betzig E 2011 Rapid three-dimensional isotropic imaging of living cells using Bessel beam plane illumination *Nat. Methods* **8** 417–23
- [79] Gao L, Shao L, Chen B-C and Betzig E 2014 3D live fluorescence imaging of cellular dynamics using Bessel beam plane illumination microscopy. *Nat. Protoc.* **9** 1083–101
- [80] Chen B-C, Legant W R, Wang K, Shao L, Milkie D E, Davidson M W, Janetopoulos C, Wu X S, Hammer J A, Liu Z, English B P, Mimori-Kiyosue Y, Romero D P, Ritter A T, Lippincott-Schwartz J, Fritz-Laylin L, Mullins R D, Mitchell D M, Bembenek J N, Reymann A-C, Bohme R, Grill S W, Wang J T, Seydoux G, Tulu U S, Kiehart D P and Betzig E 2014 Lattice light-sheet microscopy: Imaging molecules to embryos at high spatiotemporal resolution *Science (80-.)*. **346** 1257998–1257998
- [81] Vettenburg T, Dalgarno H I C, Nytk J, Coll-Lladó C, Ferrier D E K, Čížmár T, Gunn-Moore F J, Dholakia K, Coll-Llado C, Ferrier D E K, Cizmar T, Gunn-Moore F J and Dholakia K 2014 Light-sheet microscopy using an Airy beam *Nat. Methods* **11** 541–4
- [82] Yang Z, Prokopas M, Nytk J, Coll-Lladó C, Gunn-Moore F J, Ferrier D E K, Vettenburg T and Dholakia K 2014 A compact Airy beam light sheet microscope with a tilted cylindrical lens. *Biomed. Opt. Express* **5** 3434–42

- 1
 - 2
 - 3
 - 4
 - 5
 - 6
 - 7
 - 8
 - 9
 - 10
 - 11
 - 12
 - 13
 - 14
 - 15
 - 16
 - 17
 - 18
 - 19
 - 20
 - 21
 - 22
 - 23
 - 24
 - 25
 - 26
 - 27
 - 28
 - 29
 - 30
 - 31
 - 32
 - 33
 - 34
 - 35
 - 36
 - 37
 - 38
 - 39
 - 40
 - 41
 - 42
 - 43
 - 44
 - 45
 - 46
 - 47
 - 48
 - 49
 - 50
 - 51
 - 52
 - 53
 - 54
 - 55
 - 56
 - 57
 - 58
 - 59
 - 60
- [83] Galland R, Greci G, Aravind A, Viasnoff V, Studer V and Sibarita J-B B 2015 3D high- and super-resolution imaging using single-objective SPIM *Nat. Methods* **12** 641–4
- [84] Meddens M B M, Liu S, Finnegan P S, Edwards T L, James C D and Lidke K A 2016 Single objective light-sheet microscopy for high-speed whole-cell 3D super-resolution *Biomed. Opt. Express* **7** 2219
- [85] Breuninger T, Greger K and Stelzer E H K 2007 Lateral modulation boosts image quality in single plane illumination fluorescence microscopy *Opt. Lett.* **32** 1938
- [86] Schröter T J, Johnson S B, John K, Santi P a., Schroter T J, Johnson S B, John K and Santi P a. 2012 Scanning thin-sheet laser imaging microscopy (sTSLIM) with structured illumination and HiLo background rejection *Biomed. Opt. Express* **3** 170
- [87] Hoda Moosavi S, Gohn-Kreuz C, Rohrbach A, Moosavi S H, Gohn-Kreuz C and Rohrbach A 2013 Feedback phase correction of Bessel beams in confocal line light-sheet microscopy: a simulation study *Appl. Opt.* **52** 5835
- [88] Nylk J, Mitchell C, Vettenburg T, Gunn-Moore F J and Dholakia K 2014 Wavefront shaping of a Bessel light field enhances light sheet microscopy with scattered light *Three-Dimensional and Multidimensional Microscopy: Image Acquisition and Processing Xxi* vol 8949p 89490V
- [89] Gao X, Wang Q, Yun M, Guo H, Wang J and Zhuang S 2011 Focal shift of cylindrical vector axisymmetric Bessel-modulated Gaussian beam with radial variance phase wavefront *Opt. - Int. J. Light Electron Opt.* **122** 2124–8
- [90] Fahrbach F O, Gurchenkov V, Alessandri K, Nassoy P and Rohrbach A 2013 Light-sheet microscopy in thick media using scanned Bessel beams and two-photon fluorescence excitation *Opt. Express* **21** 13824
- [91] Fahrbach F O, Gurchenkov V, Alessandri K, Nassoy P and Rohrbach A 2013 Self-reconstructing sectioned Bessel beams offer submicron optical sectioning for large fields of view in light-sheet microscopy *Opt. Express* **21** 11425–40
- [92] Nylk J, McCluskey K, Aggarwal S, Tello J A and Dholakia K 2016 Enhancement of image quality and imaging depth with Airy light-sheet microscopy in cleared and non-cleared neural tissue *Biomed. Opt. Express* **7** 4021
- [93] Golub J, Chebbi B and Golub I 2015 Reducing the divergence of a light sheet below diffraction limit *2015 Photonics North (IEEE)* pp 1–1
- [94] Ebeling C G and Jorgensen E M 2013 Two views on light sheets *Nat. Biotechnol.* **31** 992–3
- [95] Wu Y, Wawrzusin P, Senseney J, Fischer R S, Christensen R, Santella A, York A G, Winter P W, Waterman C M, Bao Z, Colón-Ramos D A, McAuliffe M and Shroff H 2013 Spatially isotropic four-dimensional imaging with dual-view plane illumination microscopy *Nat. Biotechnol.* **31** 1032–8
- [96] Buytaert J a N and Dirckx J J J 2011 Design and quantitative resolution measurements of an optical virtual sectioning three-dimensional imaging technique for biomedical specimens, featuring two-micrometer slicing resolution. *J. Biomed. Opt.* **12** 14039
- [97] Manton J D and Rees E J 2016 triSPIM: light sheet microscopy with isotropic super-resolution *Opt. Lett.* **41** 4170
- [98] Dean K M, Roudot P, Welf E S, Danuser G and Fiolka R 2015 Deconvolution-free Subcellular Imaging with Axially Swept Light Sheet Microscopy *Biophys. J.* **108** 2807–15
- [99] Dean K M, Roudot P, Reis C R, Welf E S, Mettlen M and Fiolka R 2016 Diagonally Scanned Light-Sheet Microscopy for Fast Volumetric Imaging of Adherent Cells *Biophys. J.* **110** 1456–65
- [100] Theer P, Dragneva D and Knop M 2016 pi SPIM: high NA high resolution isotropic light-sheet imaging in cell culture dishes *Sci. Rep.* **6** 32880
- [101] McGorty R, Liu H, Kamiyama D, Dong Z, Guo S and Huang B 2015 Open-top selective plane illumination microscope for conventionally mounted specimens *Opt. Express* **23** 16142

- 1
 - 2
 - 3
 - 4
 - 5
 - 6
 - 7
 - 8
 - 9
 - 10
 - 11
 - 12
 - 13
 - 14
 - 15
 - 16
 - 17
 - 18
 - 19
 - 20
 - 21
 - 22
 - 23
 - 24
 - 25
 - 26
 - 27
 - 28
 - 29
 - 30
 - 31
 - 32
 - 33
 - 34
 - 35
 - 36
 - 37
 - 38
 - 39
 - 40
 - 41
 - 42
 - 43
 - 44
 - 45
 - 46
 - 47
 - 48
 - 49
 - 50
 - 51
 - 52
 - 53
 - 54
 - 55
 - 56
 - 57
 - 58
 - 59
 - 60
- [102] Wu Y, Ghitani A, Christensen R, Santella A, Du Z, Rondeau G, Bao Z, Colón-Ramos D, Shroff H, Colon-Ramos D and Shroff H 2011 Inverted selective plane illumination microscopy (iSPIM) enables coupled cell identity lineaging and neurodevelopmental imaging in *Caenorhabditis elegans*. *Proceedings of the National Academy of Sciences of the United States of America* vol 108pp 17708–13
- [103] Yang Z, Haslehurst P, Scott S, Emptage N and Dholakia K 2016 A compact light-sheet microscope for the study of the mammalian central nervous system *Sci. Rep.* **6** 26317
- [104] Capoulade J, Wachsmuth M, Hufnagel L and Knop M 2011 Quantitative fluorescence imaging of protein diffusion and interaction in living cells *Nat. Biotechnol.* **29** 835–9
- [105] Tokunaga M, Imamoto N and Sakata-Sogawa K 2008 Highly inclined thin illumination enables clear single-molecule imaging in cells *Nat. Methods* **5** 159–61
- [106] Cutrale F and Gratton E 2012 Inclined selective plane illumination microscopy adaptor for conventional microscopes *Microsc. Res. Tech.* **75** 1461–6
- [107] Hu Y S, Zhu Q, Elkins K, Tse K, Li Y, Fitzpatrick J a J, Verma I M and Cang H 2013 Light-sheet Bayesian microscopy enables deep-cell super-resolution imaging of heterochromatin in live human embryonic stem cells *Opt. Nanoscopy* **2** 7
- [108] Dunsby C 2008 Optically sectioned imaging by oblique plane microscopy. *Opt. Express* **16** 20306
- [109] Gebhardt J C M, Suter D M, Roy R, Zhao Z W, Chapman A R, Basu S, Maniatis T and Xie X S 2013 Single-molecule imaging of transcription factor binding to DNA in live mammalian cells *Nat. Methods* **10** 421–6
- [110] Greiss F, Deligiannaki M, Jung C, Gaul U and Braun D 2016 Single-Molecule Imaging in Living *Drosophila* Embryos with Reflected Light-Sheet Microscopy *Biophys. J.* **110** 939–46
- [111] Li T, Ota S, Kim J, Wong Z J, Wang Y, Yin X and Zhang X 2014 Axial Plane Optical Microscopy *Sci. Rep.* **4** 7253
- [112] Bouchard M B, Voleti V, Mendes C S, Lacefield C, Grueber W B, Mann R S, Bruno R M and Hillman E M C 2015 Swept confocally-aligned planar excitation (SCAPE) microscopy for high speed volumetric imaging of behaving organisms. *Nat. Photonics* **9** 113–9
- [113] Mitchell T J, Saunter C D, O’Nions W, Girkin J M and Love G D 2014 Quantitative high dynamic range beam profiling for fluorescence microscopy *Rev. Sci. Instrum.* **85** 103713
- [114] Wilding D, Pozzi P, Soloviev O, Vdovin G and Verhaegen M 2016 Adaptive illumination based on direct wavefront sensing in a light-sheet fluorescence microscope *Opt. Express* **24** 24896
- [115] Tomer R, Lovett-Barron M, Kauvar I, Andalman A, Burns V M, Sankaran S, Grosenick L, Buxton M, Yang S and Deisseroth K 2015 SPED Light Sheet Microscopy: Fast Mapping of Biological System Structure and Function *Cell* **163** 1796–806
- [116] Supatto W, Truong T V., Débarre D, Beaurepaire E, D?barre D, Beaurepaire E, Débarre D and Beaurepaire E 2011 Advances in multiphoton microscopy for imaging embryos *Curr. Opin. Genet. Dev.* **21** 538–48
- [117] Zong W, Zhao J, Chen X, Lin Y, Ren H, Zhang Y, Fan M, Zhou Z, Cheng H, Sun Y and Chen L 2015 Large-field high-resolution two-photon digital scanned light-sheet microscopy *Cell Res.* **25** 254–7
- [118] Lavagnino Z, Zanicchi F C, Ronzitti E and Diaspro A 2013 Two-photon excitation selective plane illumination microscopy (2PE-SPIM) of highly scattering samples: characterization and application. *Opt. Express* **21** 5998–6008
- [119] Mahou P, Vermot J, Beaurepaire E and Supatto W 2014 Multicolor two-photon light-sheet microscopy *Nat. Methods* **11** 600–1
- [120] Zhao M, Zhang H, Li Y, Ashok A, Liang R, Zhou W and Peng L 2014 Cellular imaging of deep organ using two-photon Bessel light-sheet nonlinear structured illumination microscopy *Biomed. Opt. Express* **5** 1296

- 1
 - 2
 - 3
 - 4
 - 5
 - 6
 - 7
 - 8
 - 9
 - 10
 - 11
 - 12
 - 13
 - 14
 - 15
 - 16
 - 17
 - 18
 - 19
 - 20
 - 21
 - 22
 - 23
 - 24
 - 25
 - 26
 - 27
 - 28
 - 29
 - 30
 - 31
 - 32
 - 33
 - 34
 - 35
 - 36
 - 37
 - 38
 - 39
 - 40
 - 41
 - 42
 - 43
 - 44
 - 45
 - 46
 - 47
 - 48
 - 49
 - 50
 - 51
 - 52
 - 53
 - 54
 - 55
 - 56
 - 57
 - 58
 - 59
 - 60
- [121] Chen X, Zong W, Li R, Zeng Z, Zhao J, Xi P, Chen L and Sun Y 2016 Two-photon light-sheet nanoscopy by fluorescence fluctuation correlation analysis *Nanoscale* **8** 9982–7
- [122] Maruyama A, Oshima Y, Kajiura-Kobayashi H, Nonaka S, Imamura T and Naruse K 2014 Wide field intravital imaging by two-photon-excitation digital-scanned light-sheet microscopy (2p-DSLM) with a high-pulse energy laser *Biomed. Opt. Express* **5** 3311–25
- [123] Lavagnino Z, Sancataldo G, d'Amora M, Follert P, De Pietri Tonelli D, Diaspro A and Cella Zanacchi F 2016 4D (x-y-z-t) imaging of thick biological samples by means of Two-Photon inverted Selective Plane Illumination Microscopy (2PE-iSPIM) *Sci. Rep.* **6** 23923
- [124] Palero J, Santos S I C O C O, Artigas D and Loza-Alvarez P 2010 A simple scanless two-photon fluorescence microscope using selective plane illumination *Opt. Express* **18** 8491
- [125] Truong T V, Supatto W, Koos D S, Choi J M and Fraser S E 2011 Deep and fast live imaging with two-photon scanned light-sheet microscopy. *Nat. Methods* **8** 757–60
- [126] Winter P W, York A G, Nogare D D, Ingaramo M, Christensen R, Chitnis A, Patterson G H and Shroff H 2014 Two-photon instant structured illumination microscopy improves the depth penetration of super-resolution imaging in thick scattering samples. *Optica* **1** 181–91
- [127] Barman I, Tan K M and Singh G P 2010 Optical sectioning using single-plane-illumination Raman imaging *J. Raman Spectrosc.* **41** 1099–101
- [128] Oshima Y, Sato H, Kajiura-Kobayashi H, Kimura T, Naruse K and Nonaka S 2012 Light sheet-excited spontaneous Raman imaging of a living fish by optical sectioning in a wide field Raman microscope *Opt. Express* **20** 16195
- [129] Rocha-Mendoza I, Licea-rodriguez J, Marro M, Olarte O E, Plata-Sanchez M and Loza-Alvarez P 2015 Rapid spontaneous Raman light sheet microscopy using cw-lasers and tunable filters *Biomed. Opt. Express* **6** 3449–61
- [130] Krieger J W, Singh A P, Garbe C S, Wohland T and Langowski J 2014 Dual-color fluorescence cross-correlation spectroscopy on a single plane illumination microscope (SPIM-FCCS) *Opt Express* **22** 2358–75
- [131] Struntz P and Weiss M 2016 Multiplexed measurement of protein diffusion in *Caenorhabditis elegans* embryos with SPIM-FCS *J. Phys. D-Applied Phys.* **49**
- [132] Greger K, Neetz M J, Reynaud E G and Stelzer E H K 2011 Three-dimensional Fluorescence Lifetime Imaging with a Single Plane Illumination Microscope provides an improved signal to noise ratio. *Opt. Express* **19** 20743–50
- [133] Jahr W, Schmid B, Schmied C, Fahrbach F O and Huisken J 2015 Hyperspectral light sheet microscopy *Nat. Commun.* **6** 7990
- [134] Wulstein D M, Regan K E, Robertson-Anderson R M and McGorty R 2016 Light-sheet microscopy with digital Fourier analysis measures transport properties over large field-of-view *Opt. Express* **24** 20881
- [135] Yang Z, Piksarv P, Ferrier D E K, Gunn-Moore F J and Dholakia K 2015 Macro-optical trapping for sample confinement in light sheet microscopy *Biomed. Opt. Express* **6** 2778
- [136] Cella Zanacchi F, Lavagnino Z, Perrone Donnorso M, Del Bue A, Furia L, Faretta M and Diaspro A 2011 Live-cell 3D super-resolution imaging in thick biological samples *Nat. Methods* **8** 1047–9
- [137] Cella Zanacchi F, Lavagnino Z, Faretta M, Furia L, Diaspro A, Zanacchi F C, Lavagnino Z, Faretta M, Furia L and Diaspro A 2013 Light-Sheet Confined Super-Resolution Using Two-Photon Photoactivation ed C M Aegerter *PLoS One* **8** e67667
- [138] Hu Y S, Zimmerley M, Li Y, Watters R and Cang H 2014 Single-Molecule Super-Resolution Light-Sheet Microscopy *ChemPhysChem* **15** 577–86
- [139] Kaplan C and Ewers H 2015 Optimized sample preparation for single-molecule localization-based superresolution microscopy in yeast *Nat. Protoc.* **10** 1007–21
- [140] Friedrich M, Nozadze R, de Keijzer S, Steinmeyer R, Ermolayev V and Harms G S

- 2011 Detection of Single Quantum Dots in Model Systems with Sheet Illumination Microscopy *J. Fluoresc.* 1–11
- [141] Friedrich M, Nozadze R, Gan Q, Zelman-Femiak M, Ermolayev V, Wagner T U and Harms G S 2009 Detection of single quantum dots in model organisms with sheet illumination microscopy *Biochem. Biophys. Res. Commun.* **390** 722–7
- [142] Ritter J G, Veith R, Siebrasse J-P and Kubitscheck U 2008 High-contrast single-particle tracking by selective focal plane illumination microscopy. *Opt. Express* **16** 7142–52
- [143] Friedrich M, Gan Q, Ermolayev V and Harms G S 2011 STED-SPIM: Stimulated emission depletion improves sheet illumination microscopy resolution *Biophys. J.* **100** L43-5
- [144] Friedrich M and Harms G S 2015 Axial resolution beyond the diffraction limit of a sheet illumination microscope with stimulated emission depletion *J. Biomed. Opt.* **20** 106006
- [145] Gohn-Kreuz C and Rohrbach A 2016 Light-sheet generation in inhomogeneous media using self-reconstructing beams and the STED-principle *Opt. Express* **24** 5855–65
- [146] Scheul T, Wang I I and Vial J-C 2014 STED-SPIM made simple *Opt. Express* **22** 30852–64
- [147] Hoyer P, de Medeiros G, Balázs B, Norlin N, Besir C, Hanne J, Kräusslich H-G, Engelhardt J, Sahl S J, Hell S W and Hufnagel L 2016 Breaking the diffraction limit of light-sheet fluorescence microscopy by RESOLFT *Proc. Natl. Acad. Sci.* **113** 3442–6
- [148] Kromm D, Thumberger T and Wittbrodt J 2016 An eye on light-sheet microscopy *Methods in Cell Biology* vol 133pp 105–23
- [149] Huiskens J and Stainier D Y R 2009 Selective plane illumination microscopy techniques in developmental biology *Development* **136** 1963–75
- [150] Patterson G H 2009 Fluorescence microscopy below the diffraction limit *Semin. Cell Dev. Biol.* **20** 886–93
- [151] Power R M and Huiskens J 2017 A guide to light-sheet fluorescence microscopy for multiscale imaging *Nat. Methods* **14** 360–73
- [152] Maizel A, von Wangenheim D, Federici F F, Haseloff J and Stelzer E H K K 2011 High-resolution live imaging of plant growth in near physiological bright conditions using light sheet fluorescence microscopy *Plant J.* **68** 377–85
- [153] Kaufmann A, Mickoleit M, Weber M and Huiskens J 2012 Multilayer mounting enables long-term imaging of zebrafish development in a light sheet microscope *Development* **139** 3242–7
- [154] Taylor J M, Saunter C D, Love G D, Girkin J M, Henderson D J and Chaudhry B 2011 Real-time optical gating for three-dimensional beating heart imaging *J. Biomed. Opt.* **16** 116021
- [155] Lee J, Fei P, Packard R R S, Kang H, Xu H, Baek K I, Jen N, Chen J, Yen H, Kuo C-C J, Chi N C, Ho C-M, Li R and Hsiai T K 2016 4-Dimensional light-sheet microscopy to elucidate shear stress modulation of cardiac trabeculation *J. Clin. Invest.* **126** 1679–90
- [156] Engelbrecht C J and Stelzer E H K 2006 Resolution enhancement in a light-sheet-based microscope (SPIM). *Opt. Lett.* **31** 1477–9
- [157] Oshima Y, Kajiura-Kobayashi H and Nonaka S 2012 Multimodal light-sheet microscopy for fluorescence live imaging *Three-Dimensional and Multidimensional Microscopy: Image Acquisition and Processing Xix* vol 8227, ed J A Conchello, C J Cogswell, T Wilson and T G Brown p 82271H
- [158] Komis G, Samajova O, Ovecka M, Samaj J, Šamajová O, Ovečka M and Šamaj J 2015 Super-resolution Microscopy in Plant Cell Imaging *Trends Plant Sci.* **20** 834–43
- [159] Bulina M E, Lukyanov K A, Britanova O V, Onichtchouk D, Lukyanov S and Chudakov D M 2006 Chromophore-assisted light inactivation (CALI) using the phototoxic fluorescent protein KillerRed *Nat. Protoc.* **1** 947–53
- [160] Buckley C, Carvalho M T, Young L K, Rider S A, McFadden C, Berlage C, Verdon R

- F, Taylor J M, Girkin J M and Mullins J J 2017 Precise spatio-temporal control of rapid optogenetic cell ablation with mem-KillerRed in Zebrafish *Sci. Rep.* **7** 5096
- [161] Abe J, Ozga A J A J, Swoger J, Sharpe J, Ripoll J and Stein J V J V 2016 Light sheet fluorescence microscopy for in situ cell interaction analysis in mouse lymph nodes *J. Immunol. Methods* **431** 1–10
- [162] Deschout H, Raemdonck K, Stremersch S, Maoddi P, Mernier G, Renaud P, Jiguet S, Hendrix A, Bracke M, Van den Broecke R, Röding M, Rudemo M, Demeester J, De Smedt S C, Strubbe F, Neyts K and Braeckmans K 2014 On-chip light sheet illumination enables diagnostic size and concentration measurements of membrane vesicles in biofluids *Nanoscale* **6** 1741–7
- [163] Paiè P, Bragheri F, Bassi A and Osellame R 2016 Selective plane illumination microscopy on a chip *Lab Chip* **16** 1556–60
- [164] Lin M, Qiao X, Liu Q, Shao C and Su X 2016 Light-sheet-based 2D light scattering cytometry for label-free characterization of senescent cells *Biomed. Opt. Express* **7** 5170
- [165] Plöschner M, Kollárová V, Dostál Z, Nylk J, Barton-Owen T, Ferrier D E K, Chmelík R, Dholakia K and Čížmár T 2016 Multimode fibre: Light-sheet microscopy at the tip of a needle *Sci. Rep.* **5** 18050
- [166] Wu Y, Chandris P, Winter P W, Kim E Y, Jaumouillé V, Kumar A, Guo M, Leung J M, Smith C, Rey-Suarez I, Liu H, Waterman C M, Ramamurthi K S, La Riviere P J and Shroff H 2016 Simultaneous multiview capture and fusion improves spatial resolution in wide-field and light-sheet microscopy *Optica* **3** 897
- [167] Eliceiri K W, Berthold M R, Goldberg I G, Ibáñez L, Manjunath B S, Martone M E, Murphy R F, Peng H, Plant A L, Roysam B, Stuurman N, Swedlow J R, Tomancak P and Carpenter A E 2012 Biological imaging software tools *Nat. Methods* **9** 697–710
- [168] Amat F, Höckendorf B, Wan Y, Lemon W C, McDole K, Keller P J, Hockendorf B, Wan Y, Lemon W C, McDole K, Keller P J, Hockendorf B, Wan Y, Lemon W C, McDole K and Keller P J 2015 Efficient processing and analysis of large-scale light-sheet microscopy data *Nat. Protoc.* **10** 1679–96
- [169] Schmied C, Steinbach P, Pietzsch T, Preibisch S and Tomancak P 2016 An automated workflow for parallel processing of large multiview SPIM recordings *Bioinformatics* **32** 1112–4